

**THERAPEUTIC AGENTS AND METHODS OF USE THEREOF FOR THE
MODULATION OF ANGIOGENESIS**

5

Related Application

This application is a continuation-in-part of U.S. patent application Serial No. 09/704,251, filed November 1, 2000, pending, the entire contents of which are hereby incorporated by reference.

10

Background of the Invention

Angiogenesis is the fundamental process by which new blood vessels are formed and is essential to a variety of normal body activities (such as reproduction, development and wound repair). Although the process is not completely understood, it is believed to involve a complex interplay of molecules which both stimulate and inhibit the growth of endothelial cells, the primary cells of the capillary blood vessels. Under normal conditions, these molecules appear to maintain the microvasculature in a quiescent state (i.e., one of no capillary growth) for prolonged periods which may last for as long as weeks or in some cases, decades. When necessary, however, (such as during wound repair), these same cells can undergo rapid proliferation and turnover within a 5 day period (Folkman, J. and Shing, Y., *Journal of Biological Chemistry*, 267(16): 10931-10934, and Folkman, J. and Klagsbrun, M. (1987) *Science*, 235: 442-447).

Although angiogenesis is a highly regulated process under normal conditions, many diseases (characterized as "angiogenic diseases") are driven by persistent unregulated angiogenesis. Otherwise stated, unregulated angiogenesis may either cause a particular disease directly or exacerbate an existing pathological condition. For example, ocular neovascularization has been implicated as the most common cause of blindness and dominates approximately 20 eye diseases. In certain existing conditions such as arthritis, newly formed capillary blood vessels invade the joints and destroy cartilage. In diabetes, new capillaries formed in the retina invade the vitreous, bleed, and cause blindness. Growth and metastasis of solid tumors are also angiogenesis-dependent (Folkman, J. (1986) *Cancer Research* 46: 467-473 and Folkman, J. (1989) *Journal of the National Cancer Institute* 82: 4-6). It has been shown, for example, that tumors which enlarge to greater than 2 mm, must obtain their own blood supply and do so by inducing the growth of new capillary blood vessels. Once these new blood vessels become embedded in the tumor, they provide a means for tumor cells to enter the circulation and metastasize to distant sites, such as the liver, lung or bone (Weidner, N., et al. (1991) *The New England Journal of Medicine* 324(1):1-8).

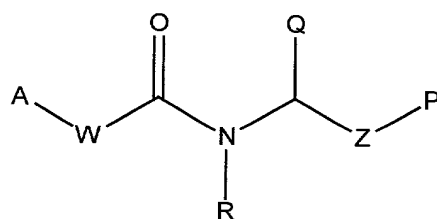
Fumagillin is a known compound which has been used as an antimicrobial and antiprotozoal. Its physicochemical properties and method of production are well known (U.S. Pat. No. 2,803,586 and *Proc. Nat. Acad. Sci. USA* (1962) 48:733-735). Fumagillin and certain types of Fumagillin analogs have also been reported to exhibit anti-angiogenic activity. However, the use of such inhibitors (e.g., TNP-470) may be limited by their rapid metabolic degradation, erratic blood levels, and by dose-limiting central nervous system (CNS) side effects.

Accordingly, there is still a need for angiogenesis inhibitors which are more potent, less neurotoxic, more stable, and/or have longer serum half-lives.

Summary of the Invention

The present invention provides angiogenesis inhibitor compounds which comprise a core, e.g., a Fumagillin core, that is believed to inhibit methionine aminopeptidase 2 (MetAP-2), coupled to a peptide. The present invention is based, at least in part, on the discovery that coupling the MetAP-2 inhibitory core to a peptide prevents the metabolic degradation of the angiogenesis inhibitor compound to ensure a superior pharmacokinetic profile and limits CNS side effects by altering the ability of the angiogenesis inhibitor compound to cross the blood brain barrier. The present invention is also based, at least in part, on the discovery that coupling the MetAP-2 inhibitory core to a peptide comprising a site-directed sequence allows for a cell specific delivery of the angiogenesis inhibitor compound and limits the toxicity of the angiogenesis inhibitor compound.

Accordingly, the present invention is directed to angiogenesis inhibitor compounds of Formula I,



(I)

where A is a MetAP-2 inhibitory core and W is O or NR. In one embodiment, Z is -C(O)- or -alkylene-C(O)- and P is NHR, OR or a peptide consisting of one to about one hundred amino acid residues connected at the N-terminus to Z. In this embodiment, Q is hydrogen, linear, branched or cyclic alkyl or aryl, provided that when P is -OR, Q is not hydrogen.

In another embodiment, Z is -alkylene-O- or -alkylene-N(R)- and P is hydrogen or a peptide consisting of from one to about one hundred amino acid residues connected to Z at the carboxyl terminus. In this embodiment, Q is hydrogen, linear, branched or cyclic alkyl or aryl, provided that when P is hydrogen, Q is not hydrogen.

5 In the angiogenesis inhibitor compounds of Formula I, each R is, independently, hydrogen or alkyl.

In another aspect, the invention features pharmaceutical compositions comprising the angiogenesis inhibitor compounds of Formula I and a pharmaceutically acceptable carrier.

10 In yet another aspect, the invention features a method of treating an angiogenic disease, *e.g.*, cancer, in a subject. The method includes administering to the subject a therapeutically effective amount of one or more angiogenesis inhibitor compounds of Formula I.

15 Other features and advantages of the invention will be apparent from the following detailed description and claims.

Brief Description of the Drawings

20 *Figure 1* is a graph depicting the results from the rat aortic ring assay (RARA) used to test the ability of the angiogenesis inhibitor compounds of the invention to inhibit angiogenesis.

Figure 2 is a graph depicting the mean daily clinical score for each of the four groups of rats tested over the 19 day period following induction of clinical arthritis.

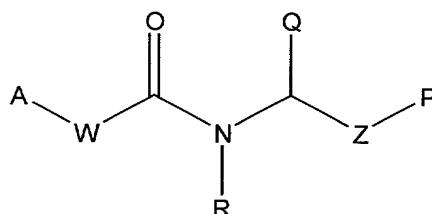
Detailed Description of the Invention

25 The present invention provides compounds useful as angiogenesis inhibitors and methods for using these compounds in the treatment of angiogenic diseases. Without intending to be limited by theory, it is believed that the angiogenesis inhibitor compounds of the invention inhibit angiogenesis by inhibiting methionine
30 aminopeptidase 2 (MetAP-2), an enzyme which cleaves the N-terminal methionine residue of newly synthesized proteins to produce the active form of the protein. At the same time, the presence of a peptide in the angiogenesis inhibitor compounds of the invention prevents the metabolic degradation of the angiogenesis inhibitor compounds and ensures a superior pharmacokinetic profile. The presence of the peptide in the
35 angiogenesis inhibitor compounds of the invention also alters the ability of the angiogenesis inhibitor compound to cross the blood brain barrier to, for example, limit CNS side effects (such as CNS toxicity). The presence of peptides comprising a site-directed sequence in the angiogenesis inhibitor compounds of the invention allows for a

site-specific delivery of the angiogenesis inhibitor compounds and, thus, limits the toxicity of the angiogenesis inhibitor compounds.

The angiogenesis inhibitor compounds of the invention comprise a MetAP-2 inhibitory core and a peptide attached, directly or indirectly, thereto. In one

embodiment, the invention provides angiogenesis inhibitor compounds of Formula I,



(I)

where A is a MetAP-2 inhibitory core and W is O or NR. In one embodiment, Z is –C(O)- or -alkylene-C(O)- and P is NHR, OR or a peptide consisting of one to about one hundred amino acid residues connected at the N-terminus to Z. In this embodiment, Q is hydrogen, linear, branched or cyclic alkyl or aryl, provided that when P is –OR, Q is not hydrogen. Z is preferably –C(O)- or C₁-C₄-alkylene-C(O)-, and, more preferably, -C(O)- or C₁-C₂-alkylene-C(O)-. Q is preferably linear, branched or cyclic C₁-C₆-alkyl, phenyl or naphthyl. More preferably, Q is isopropyl, phenyl or cyclohexyl.

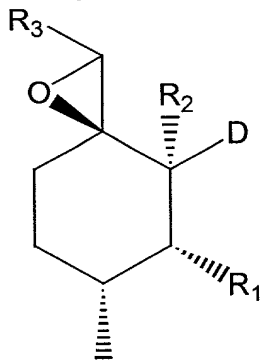
In another embodiment, Z is –alkylene-O- or –alkylene-N(R)-, where alkylene is, preferably, C₁-C₆-alkylene, more preferably C₁-C₄-alkylene and, most preferably, C₁-C₂-alkylene. P is hydrogen or a peptide consisting of from one to about one hundred amino acid residues connected to Z at the carboxyl terminus. In this embodiment, Q is hydrogen, linear, branched or cyclic alkyl or aryl, provided that when P is hydrogen, Q is not hydrogen. Q is preferably linear, branched or cyclic C₁-C₆-alkyl, phenyl or naphthyl. More preferably, Q is isopropyl, phenyl or cyclohexyl.

In the compounds of Formula I, each R is, independently, hydrogen or alkyl. In one embodiment, each R is, independently, hydrogen or linear, branched or cyclic C₁-C₆-alkyl. Preferably, each R is, independently, hydrogen or linear or branched C₁-C₄-alkyl. More preferably, each R is, independently, hydrogen or methyl. In the most preferred embodiments, each R is hydrogen.

In Formula I, A is a MetAP-2 inhibitory core. As used herein, a “MetAP-2 inhibitory core” includes a moiety able to inhibit the activity of methionine

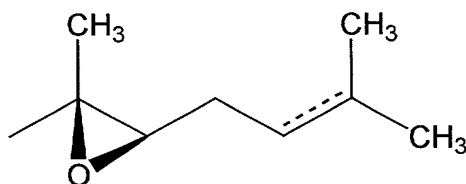
aminopeptidase 2 (MetAP-2), *e.g.*, the ability of MetAP-2 to cleave the N-terminal methionine residue of newly synthesized proteins to produce the active form of the protein. Preferred MetAP-2 inhibitory cores are Fumagillin derived structures.

Suitable MetAP-2 inhibitory cores include the cores of Formula II,



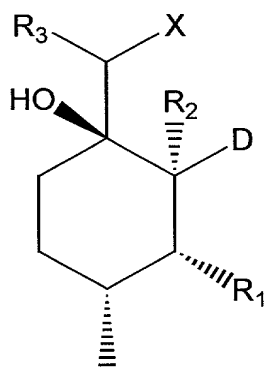
(II)

where R₁ is hydrogen or alkoxy, preferably C₁-C₄-alkoxy and more preferably, methoxy. R₂ is hydrogen or hydroxy; and R₃ is hydrogen or alkyl, preferably C₁-C₄-alkyl and more preferably, methyl. D is linear or branched alkyl, preferably C₁-C₆-alkyl; arylalkyl, preferably aryl-C₁-C₄-alkyl and more preferably phenyl-C₁-C₄-alkyl; or D is of the structure



where the dashed line represents a single bond or a double bond.

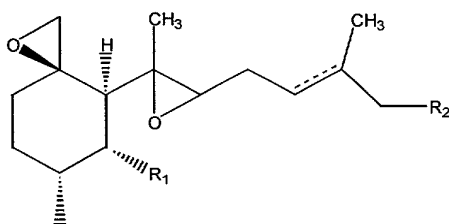
A can also be a MetAP-2 inhibitory core of Formula III,



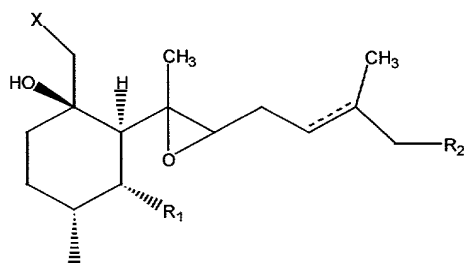
(III)

Where R_1 , R_2 , R_3 and D have the meanings given above for Formula II, and X is a leaving group, such as a halogen.

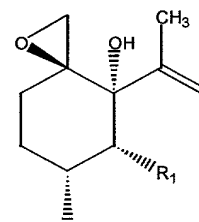
- 5 Examples of suitable MetAP-2 inhibitory cores include, but are not limited to, the following.



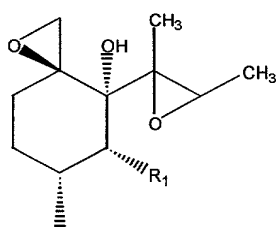
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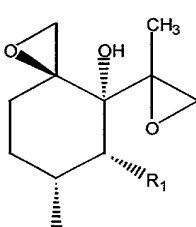
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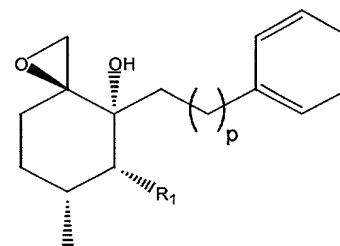
(VI)



(VII)



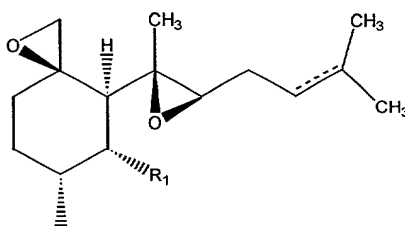
(VIII)



(IX)

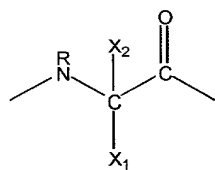
In each of Formulas IV-IX, the indicated valence on the ring carbon is the point of attachment of the linker group. In Formula IX, p is an integer from 0 to 10, preferably 1-4. In Formulas IV, V and VI-IX, R₁ is hydrogen or C₁-C₄-alkoxy, preferably methoxy. In Formulas IV and V, the dashed line indicates that the bond can be a double bond or a single bond. In Formula V, X represents a leaving group, such as a thioalkoxy group, a thioaryloxy group, a halogen or a dialkylsulfonium group. In Formulas IV and V, R₂ is H, OH, amino, C₁-C₄-alkylamino or di(C₁-C₄-alkyl)amino, preferably H. In formulas in which the stereochemistry of a particular stereocenter is not indicated, that stereocenter can have either of the possible stereochemistries, consistent with the ability of the angiogenesis inhibitor compound to inhibit the activity of MetAP-2.

In particularly preferred embodiments, A is the MetAP-2 inhibitory core of Formula X below.

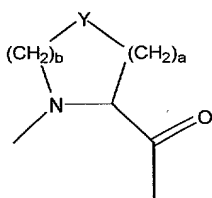


(X)

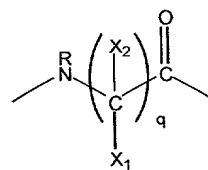
As used herein, the terms "P" and "peptide" include compounds comprising from 1 to about 100 amino acid residues (*e.g.*, 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20 or more amino acid residues). In preferred embodiments, the peptide includes compounds comprising less than about 90, 80, 70, 60, 50, 40, 30, 20, or 10 amino acid residues, preferably about 1-10, 1-20, 1-30, 1-40, 1-50, 1-60, 1-70, 1-80, or 1-90 amino acid residues. The peptides may be natural or synthetically made. The amino acid residues are preferably α -amino acid residues. For example, the amino acid residues can be independently selected from among the twenty naturally occurring amino acid residues, the D-enantiomers of the twenty natural amino acid residues, and may also be non-natural amino acid residues (*e.g.*, norleucine, norvaline, phenylglycine, β -alanine, or a peptide mimetic such as 3-amino-methylbenzoic acid). In one embodiment, the amino acid residues are independently selected from residues of Formula XI, Formula XII, and Formula XIII.



XI



XII



XIII

5

In Formula XI, X_1 is hydrogen, a side chain of one of the twenty naturally-occurring amino acid residues, a linear, branched or cyclic C_1 - C_8 -alkyl group, an aryl group, such as a phenyl or naphthyl group, an aryl- C_1 - C_4 -alkyl group, a heteroaryl group, such as a pyridyl, thienyl, pyrrolyl, or furyl group, or a heteroaryl- C_1 - C_4 -alkyl group; and X_2 is hydrogen a linear, branched or cyclic C_1 - C_8 -alkyl group, an aryl group, such as a phenyl or naphthyl group, an aryl- C_1 - C_4 -alkyl group or a heteroaryl group as described above for X_1 . Preferably, X_2 is hydrogen. In Formula XII, Y is methylene, oxygen, sulfur or NH, and a and b are each, independently, 0-4, provided that the sum of a and b is between 1 and 4. Formulas XI and XII encompass α -amino acid residues

having either a D or an L stereochemistry at the alpha carbon atom. One or more of the amino acid residues can also be an amino acid residue other than an α -amino acid residue, such as a β -, γ - or ϵ -amino acid residue. Suitable examples of such amino acid residues are of Formula XIII, wherein q is an integer of from 2 to about 6, and each X_1 and X_2 independently have the meanings given above for these variables in Formula XI.

In a preferred embodiment, the peptide used in the angiogenesis inhibitor compounds of the invention may include a site-directed sequence in order to increase the specificity of binding of the angiogenesis inhibitor compound to a cell surface of interest. As used herein, the term "site-directed sequence" is intended to include any amino acid sequence (*e.g.*, comprised of natural or non natural amino acid residues) which serves to limit exposure of the angiogenesis inhibitor compound to the periphery and/or which serves to direct the angiogenesis inhibitor compound to a site of interest, *e.g.*, a site of angiogenesis or aberrant cellular proliferation.

The peptide contained within the angiogenesis inhibitor compounds of the invention may include a peptide cleavage site for an enzyme which is expressed at sites of angiogenesis or aberrant cell proliferation, allowing tissue-selective delivery of a cell-permeable active angiogenesis inhibitor compound or fragment thereof (*e.g.*, a fragment containing the MetAP-2 inhibitory core of the angiogenesis inhibitor compound). The peptide may also include a sequence which is a ligand for a cell surface receptor which is expressed at a site of angiogenesis or aberrant cell proliferation, thereby targeting

angiogenesis inhibitor compounds to a cell surface of interest. For example, a peptide contained within the angiogenesis inhibitor compounds of the invention can include a cleavage site for a matrix metalloproteinase, or an integrin binding RGD (Arg-Gly-Asp) sequence, or a combination of both an enzyme "cleavage" sequence and a cell surface

5 "ligand" which serve to target the angiogenesis inhibitor compound to the membrane of an endothelial cell. However, the selection of a peptide sequence must be such that the active angiogenesis inhibitor compound is available to be delivered to the cells in which MetAP-2 inhibition is desired.

For example, a sequence that is cleaved by a matrix metalloproteinase produces a

10 product that contains the MetAP-2 inhibitory core, a coupling group, and a peptide fragment. Sequences are selected so that the active angiogenesis inhibitor compound, *e.g.*, the active angiogenesis inhibitor compound generated by the matrix metalloproteinase cleavage, is cell permeable. Preferably, the active angiogenesis inhibitor compound does not contain a free acid after the cleavage.

15 In one embodiment, the peptide includes a cleavage site for a matrix metalloproteinase, such as matrix metalloproteinase-2 (MMP-2), MMP-1, MMP-3, MMP-7, MMP-8, MMP-9, MMP-12, MMP-13 or MMP-26. Preferably, the peptide includes a cleavage site for MMP-2 or MMP-9. For example, the peptide can comprise the sequence -Pro-Leu-Gly-Xaa- (SEQ ID NO:1), where Xaa is any naturally occurring

20 amino acid residue consistent with matrix metalloproteinase (MMP) cleavage at the Gly-Xaa bond. Xaa is preferably a hydrophobic amino acid residue, such as tryptophan, phenylalanine, methionine, leucine, isoleucine, proline, and valine.

Other suitable sequences include sequences comprising one or more of Pro-Cha-Gly-Cys(Me)-His (SEQ ID NO:2); Pro-Gln-Gly-Ile-Ala-Gly-Gln-D-Arg (SEQ ID

25 NO:3); Pro-Gln-Gly-Ile-Ala-Gly-Trp (SEQ ID NO:4); Pro-Leu-Gly-Cys(Me)-His-Ala-D-Arg (SEQ ID NO:5); Pro-Leu-Gly-Met-Trp-Ser-Arg (SEQ ID NO:35); Pro-Leu-Gly-Leu-Trp-Ala-D-Arg (SEQ ID NO:6); Pro-Leu-Ala-Leu-Trp-Ala-Arg (SEQ ID NO:7); Pro-Leu-Ala-Leu-Trp-Ala-Arg (SEQ ID NO:8); Pro-Leu-Ala-Tyr-Trp-Ala-Arg (SEQ ID NO:9); Pro-Tyr-Ala-Tyr-Trp-Met-Arg (SEQ ID NO:10); Pro-Cha-Gly-Nva-His-Ala

30 (SEQ ID NO:11); Pro-Leu-Ala-Nva (SEQ ID NO:12); Pro-Leu-Gly-Leu (SEQ ID NO:13); Pro-Leu-Gly-Ala (SEQ ID NO:14); Arg-Pro-Leu-Ala-Leu-Trp-Arg-Ser (SEQ ID NO:15); Pro-Cha-Ala-Abu-Cys(Me)-His-Ala (SEQ ID NO:16); Pro-Cha-Ala-Gly-Cys(Me)-His-Ala (SEQ ID NO:17); Pro-Lys-Pro-Gln-Gln-Phe-Phe-Gly-Leu (SEQ ID NO:18); Pro-Lys-Pro-Leu-Ala-Leu (SEQ ID NO:19); Arg-Pro-Lys-Pro-Tyr-Ala-Nva

35 Trp-Met (SEQ ID NO:20); Arg-Pro-Lys-Pro-Val-Glu-Nva-Trp-Arg (SEQ ID NO:21); Arg-Pro-Lys-Pro-Val-Glu-Nva-Trp-Arg (SEQ ID NO:22); and Arg-Pro-Lys-Pro-Leu-Ala-Nva-Trp (SEQ ID NO:23). These sequences identify the natural amino acid residues using the customary three-letter abbreviations; the following abbreviations

represent the indicated non-natural amino acids: Abu = L-a-aminobutyryl; Cha = L-cyclohexylalanine; Nva = L-norvaline.

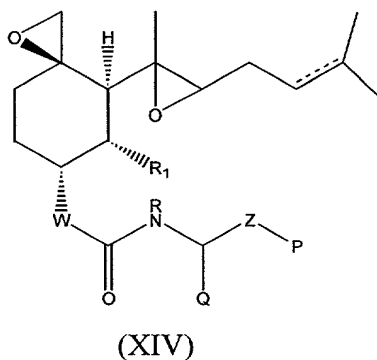
In certain embodiments, P is an amino acid sequence selected from the group consisting of Ac-Pro-Leu-Gly-Met-Trp-Ala (SEQ ID NO:24); Gly-Pro-Leu-Gly-Met-
 5 His-Ala-Gly (SEQ ID NO:25); Gly-Pro-Leu-(Me)Gly (SEQ ID NO:26); Gly-Pro-Leu-Gly (SEQ ID NO:27); Gly-Met-Gly-Leu-Pro (SEQ ID NO:28); Ala-Met-Gly-Ile-Pro (SEQ ID NO:29); Gly-Arg-Gly-Asp-(O-Me-Tyr)-Arg-Glu (SEQ ID NO:30); Gly-Arg-Gly-Asp-Ser-Pro (SEQ ID NO:31); Gly-Arg-Gly-Asp (SEQ ID NO:32); Asp-Gly-Arg; Ac-Pro-Leu-Gly-Met-Ala (SEQ ID NO:33); Ac-Arg-Gly-Asp-Ser-Pro-Leu-Gly-Met-
 10 Trp-Ala (SEQ ID NO:34); Ac-Pro-Leu-Gly-Met-Gly (SEQ ID NO:36); Met-Trp-Ala (SEQ ID NO:37); Met-Gly (SEQ ID NO:38); Gly-Pro-Leu-Gly-Met-Trp-Ala-Gly (SEQ ID NO:39); and Gly-Arg-(3-amino-3-pyriylpropionic acid) (SEQ ID NO:40). (Ac in the foregoing sequences represents an Acetyl group).

The peptide can be attached to the MetAP-2 inhibitory core at either its N-
 15 terminus or C-terminus. When the peptide is attached to the MetAP-2 inhibitory core at its C-terminus, the N-terminus of the peptide can be $-NR_2R_3$, where R_2 is hydrogen, alkyl or arylalkyl and R_3 is hydrogen, alkyl, arylalkyl or acyl. When the peptide is attached to the MetAP-2 inhibitory core at its N-terminus, the C-terminus can be $-C(O)R_4$, where R_4 is -OH, -O-alkyl, -O-arylalkyl, or $-NR_2R_3$, where R_2 is hydrogen, alkyl or arylalkyl and
 20 R_3 is hydrogen, alkyl, arylalkyl or acyl. In this embodiment, the C-terminal residue can also be present in a reduced form, such as the corresponding primary alcohol.

The present invention also includes pharmaceutically acceptable salts of the angiogenesis inhibitor compounds of the invention. A "pharmaceutically acceptable salt" includes a salt that retains the desired biological activity of the parent angiogenesis
 25 inhibitor compound and does not impart any undesired toxicological effects. Examples of such salts are salts of acids such as hydrochloric acid, hydrobromic acid, sulfuric acid, phosphoric acid, nitric acid, and the like; acetic acid, oxalic acid, tartaric acid, succinic acid, malic acid, benzoic acid, pamoic acid, alginic acid, methanesulfonic acid, naphthalenesulfonic acid, and the like. Also included are salts of cations such as sodium,
 30 potassium, lithium, zinc, copper, barium, bismuth, calcium, and the like; or organic cations such as trialkylammonium. Combinations of the above salts are also useful.

Preferred Angiogenesis Inhibitor Compounds of Formula I

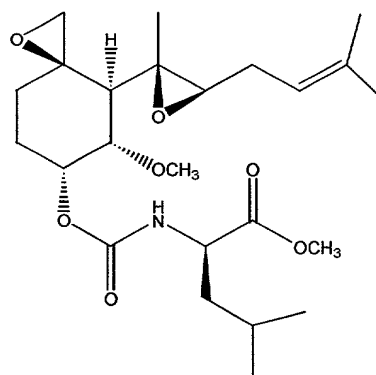
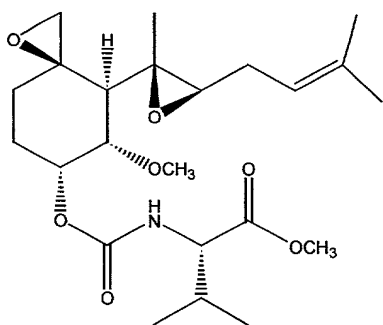
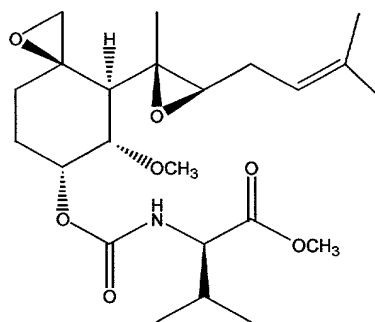
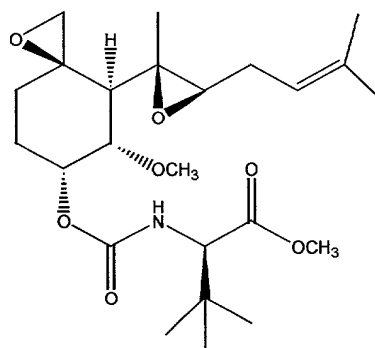
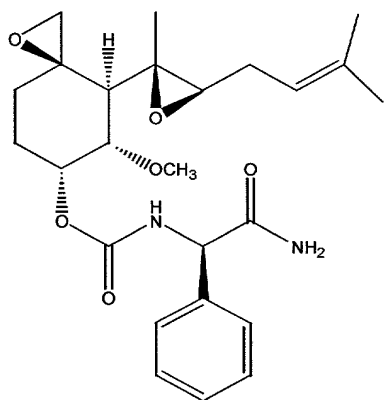
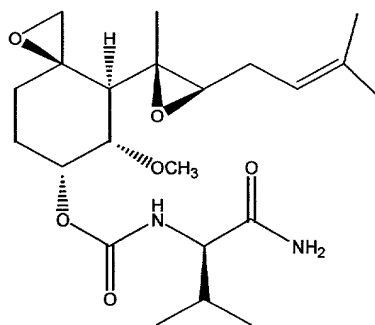
A first preferred subset of the angiogenesis inhibitor compounds of Formula I
 35 comprises Formula XIV shown below.

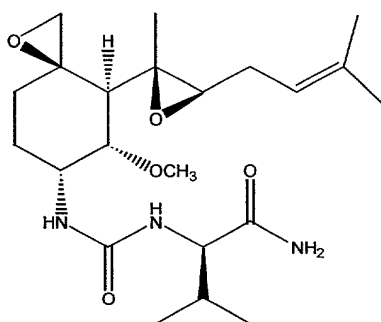
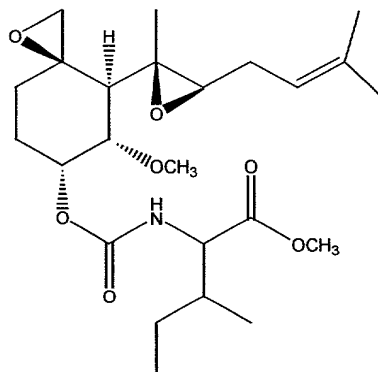
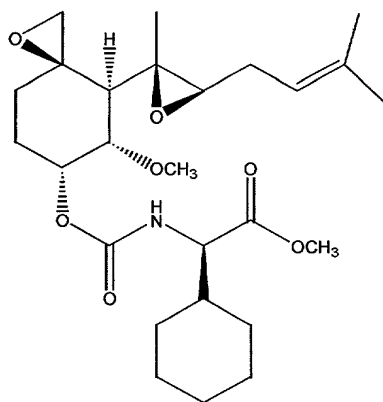


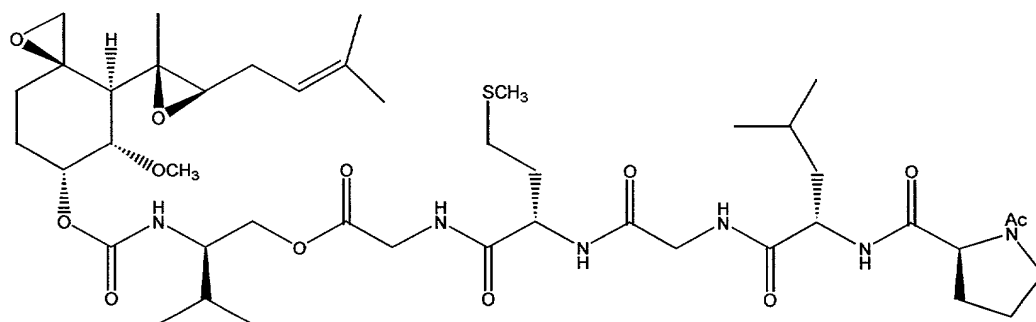
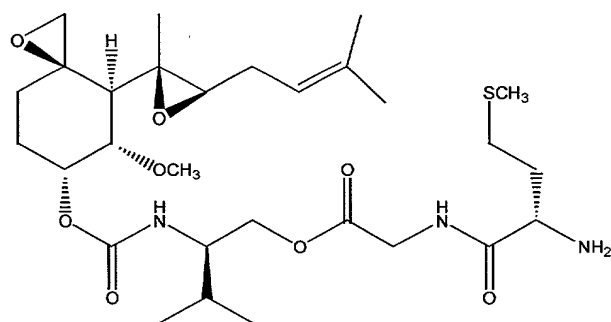
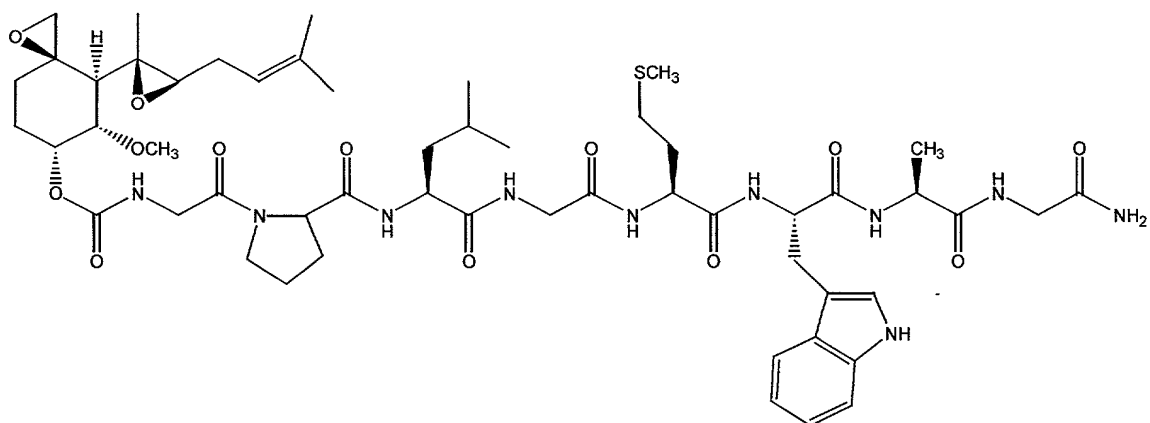
In one embodiment, W is O or NR. Z is $-C(O)$ or $-alkylene-C(O)-$, preferably C1-C4-alkylene- $C(O)-$. R is hydrogen or a C₁-C₄-alkyl. Q is hydrogen; linear, branched or cyclic C₁-C₆-alkyl; or aryl. R₁ is hydroxy, C₁-C₄-alkoxy or halogen. P is NH₂, OR or a peptide attached to Z at its N-terminus and comprising from 1 to 100 amino acid residues independently selected from naturally occurring amino acid residues, D-enantiomers of the naturally occurring amino acid residues and non-natural amino acid residues. When Q is H, P is not NH₂ or OR. In preferred embodiments, W is O or NH; Q is isopropyl; R₁ is methoxy; P comprises from 1 to 15 amino acid residues; and the dashed line present in Formula XIV represents a double bond. In particularly preferred embodiments, W is O, and P comprises 10 or fewer amino acid residues.

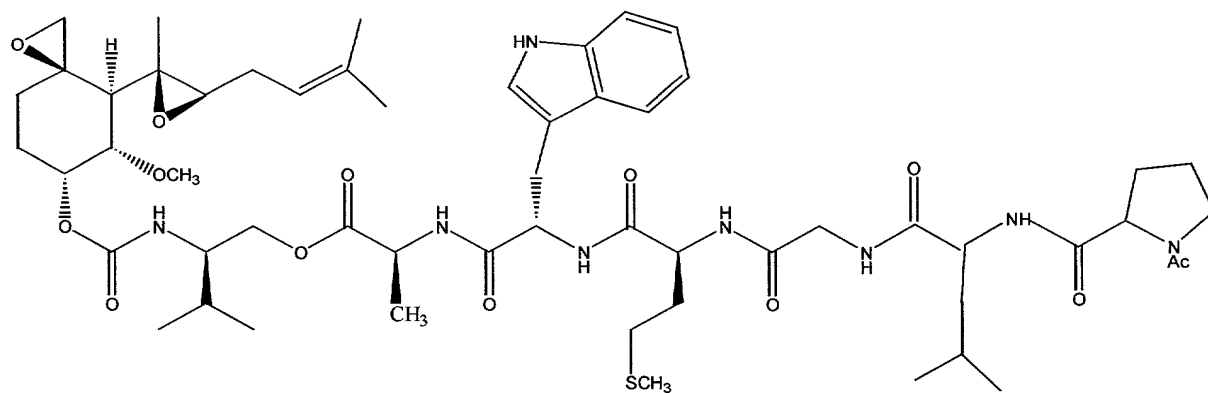
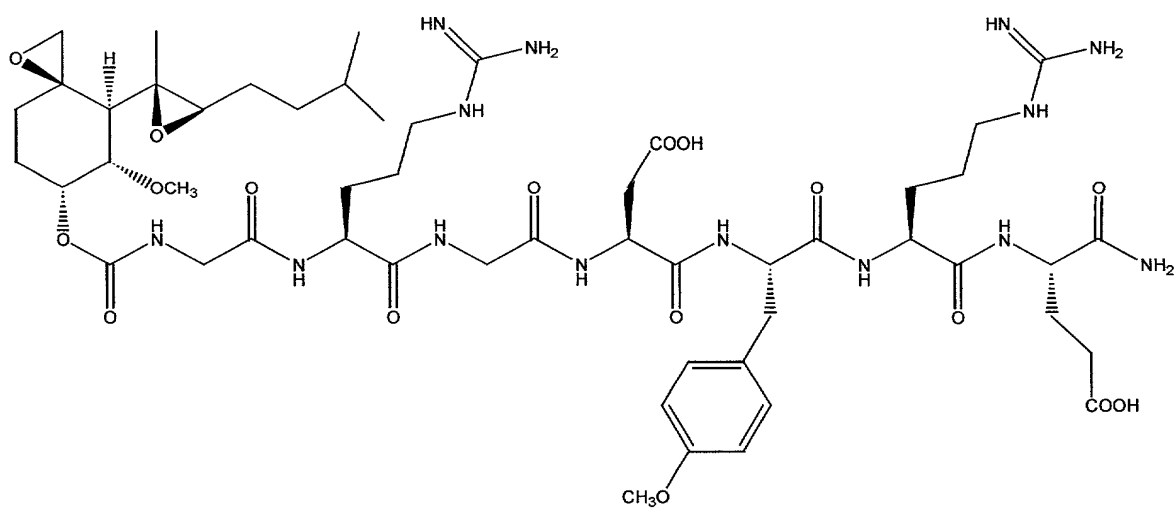
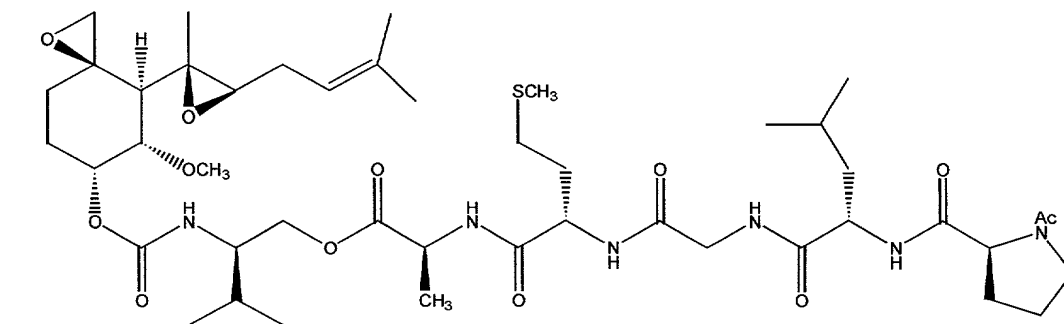
In another embodiment of the compounds of Formula XIV, W is O or NR. Z is alkylene-O or alkylene-NR, preferably C1-C4-alkylene-O or C1-C4-alkylene-NR-. R is hydrogen or a C₁-C₄-alkyl. Q is hydrogen; linear, branched or cyclic C₁-C₆-alkyl; or aryl. R₁ is hydroxy, C₁-C₄-alkoxy or halogen. P is hydrogen or a peptide attached to Z at its C-terminus and comprising from 1 to 100 amino acid residues independently selected from naturally occurring amino acid residues, D-enantiomers of the naturally occurring amino acid residues and non-natural amino acid residues. When Q is H, P is not H. In preferred embodiments, W is O or NH; Q is isopropyl; R₁ is methoxy; P comprises from 1 to 15 amino acid residues; and the dashed line present in Formula XIV represents a double bond. In particularly preferred embodiments, W is O, and P comprises 10 or fewer amino acid residues or P is hydrogen.

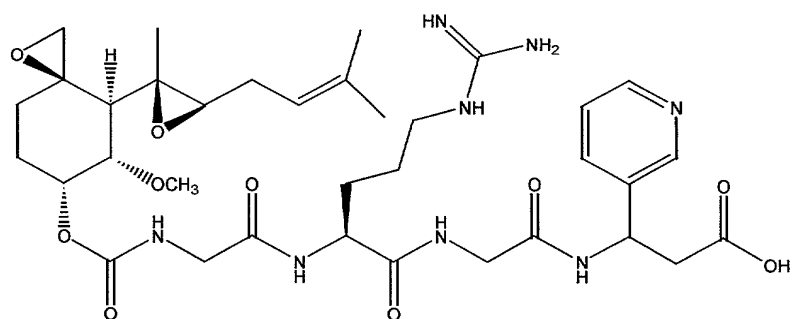
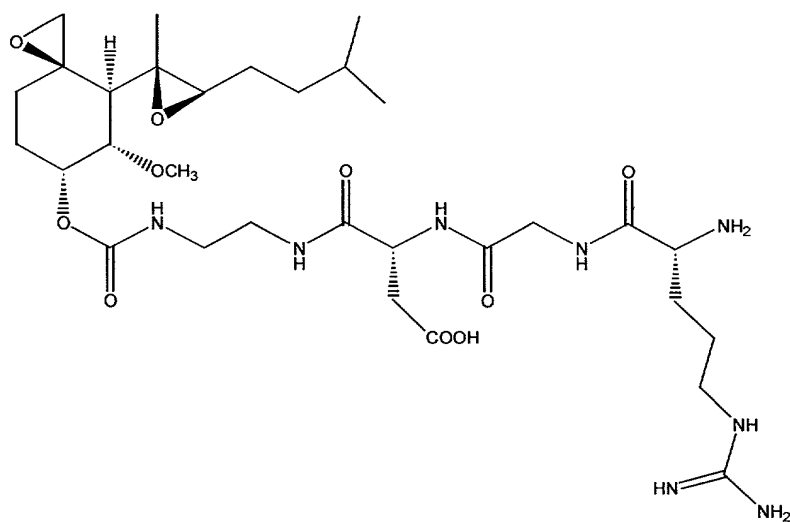
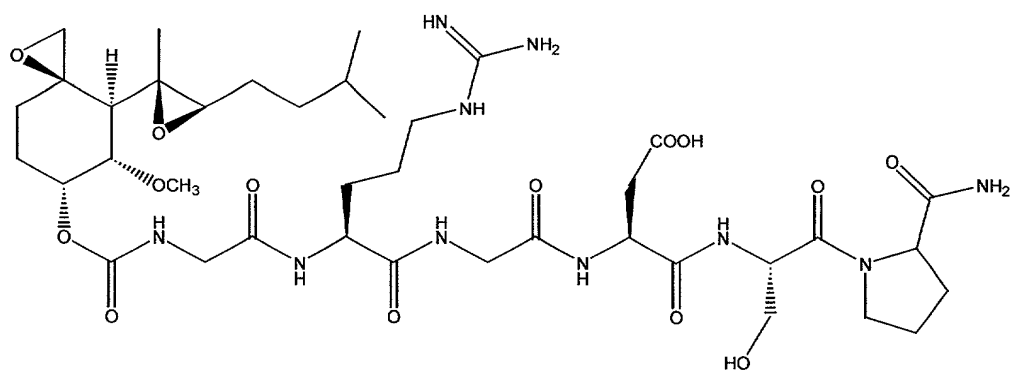
One set of particularly preferred angiogenesis inhibitor compounds of the invention is represented by the structures set forth below.

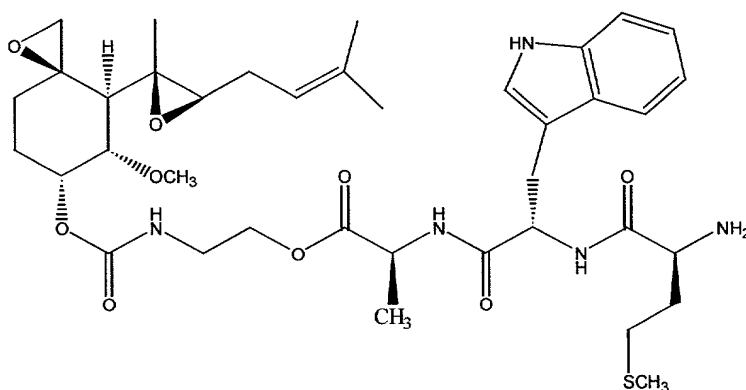
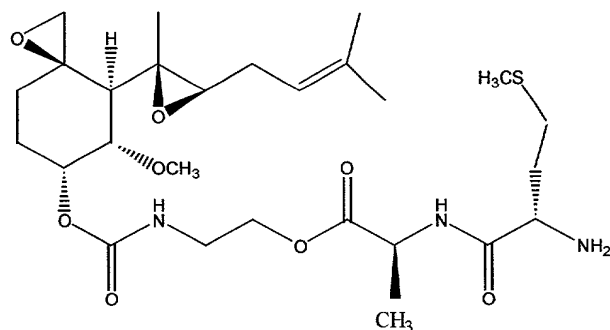












Methods of Using the Angiogenesis Inhibitor Compounds for the Treatment of Angiogenic Disease

- 5 In another embodiment, the present invention provides a method of treating an angiogenic disease in a subject. The method includes administering to the subject a therapeutically effective amount of an angiogenesis inhibitor compound of the present invention, thereby treating the angiogenic disease in the subject.

- As used herein, the term “angiogenic disease” includes a disease, disorder, or
 10 condition characterized or caused by aberrant or unwanted, *e.g.*, stimulated or suppressed, formation of blood vessels (angiogenesis). Aberrant or unwanted angiogenesis may either cause a particular disease directly or exacerbate an existing pathological condition. Examples of angiogenic diseases include ocular disorders, *e.g.*, diabetic retinopathy, retinopathy of prematurity, corneal graft rejection, retrolental
 15 fibroplasia, neovascular glaucoma, rubeosis, retinal neovascularization due to macular degeneration, hypoxia, angiogenesis in the eye associated with infection or surgical intervention, ocular tumors and trachoma, and other abnormal neovascularization conditions of the eye, where neovascularization may lead to blindness; disorders

affecting the skin, *e.g.*, psoriasis and pyogenic granuloma; cancer, *e.g.*, carcinomas and sarcomas, where progressive growth is dependent upon the continuous induction of angiogenesis by these tumor cells; pediatric disorders, *e.g.*, angiofibroma, and hemophiliac joints; blood vessel diseases such as hemangiomas, and capillary proliferation within atherosclerotic plaques; disorders associated with surgery, *e.g.*, hypertrophic scars, wound granulation and vascular adhesions; and autoimmune diseases such as rheumatoid, immune and degenerative arthritis, where new vessels in the joint may destroy articular cartilage and scleroderma.

The term angiogenic disease also includes diseases characterized by excessive or abnormal stimulation of endothelial cells, including but not limited to intestinal adhesions, Crohn's disease, atherosclerosis, scleroderma, and hypertrophic scars, *i.e.*, keloids; diseases that have angiogenesis as a pathologic consequence such as cat scratch disease (*Rochele ninalia quintosa*) and ulcers (*Helicobacter pylori*). In addition, the angiogenesis inhibitor compounds of the present invention are useful as birth control agents (by virtue of their ability to inhibit the angiogenesis dependent ovulation and establishment of the placenta) and may also be used to reduce bleeding by administration to a subject prior to surgery.

As used herein, the term "subject" includes warm-blooded animals, preferably mammals, including humans. In a preferred embodiment, the subject is a primate. In an even more preferred embodiment, the primate is a human.

As used herein, the term "administering" to a subject includes dispensing, delivering or applying an angiogenesis inhibitor compound, *e.g.*, an angiogenesis inhibitor compound in a pharmaceutical formulation (as described herein), to a subject by any suitable route for delivery of the compound to the desired location in the subject, including delivery by either the parenteral or oral route, intramuscular injection, subcutaneous/intradermal injection, intravenous injection, buccal administration, transdermal delivery and administration by the rectal, colonic, vaginal, intranasal or respiratory tract route.

As used herein, the term "effective amount" includes an amount effective, at dosages and for periods of time necessary, to achieve the desired result, *e.g.*, sufficient to treat an angiogenic disease in a subject. An effective amount of an angiogenesis inhibitor compound, as defined herein may vary according to factors such as the disease state, age, and weight of the subject, and the ability of the angiogenesis inhibitor compound to elicit a desired response in the subject. Dosage regimens may be adjusted to provide the optimum therapeutic response. An effective amount is also one in which any toxic or detrimental effects (*e.g.*, side effects) of the angiogenesis inhibitor compound are outweighed by the therapeutically beneficial effects.

A therapeutically effective amount of an angiogenesis inhibitor compound (*i.e.*, an effective dosage) may range from about 0.001 to 30 mg/kg body weight, preferably about 0.01 to 25 mg/kg body weight, more preferably about 0.1 to 20 mg/kg body weight, and even more preferably about 1 to 10 mg/kg, 2 to 9 mg/kg, 3 to 8 mg/kg, 4 to 7 mg/kg, or 5 to 6 mg/kg body weight. The skilled artisan will appreciate that certain factors may influence the dosage required to effectively treat a subject, including but not limited to the severity of the disease or disorder, previous treatments, the general health and/or age of the subject, and other diseases present. Moreover, treatment of a subject with a therapeutically effective amount of an angiogenesis inhibitor compound can include a single treatment or, preferably, can include a series of treatments. In one example, a subject is treated with an angiogenesis inhibitor compound in the range of between about 0.1 to 20 mg/kg body weight, one time per week for between about 1 to 10 weeks, preferably between 2 to 8 weeks, more preferably between about 3 to 7 weeks, and even more preferably for about 4, 5, or 6 weeks. It will also be appreciated that the effective dosage of an angiogenesis inhibitor compound used for treatment may increase or decrease over the course of a particular treatment.

The methods of the invention further include administering to a subject a therapeutically effective amount of an angiogenesis inhibitor compound in combination with another pharmaceutically active compound known to treat an angiogenic disease, *e.g.*, a chemotherapeutic agent such as Taxol, Paclitaxel, or Actinomycin D, or an antidiabetic agent such as Tolbutamide; or a compound that may potentiate the angiogenesis inhibitory activity of the angiogenesis inhibitor compound, such as heparin or a sulfated cyclodextrin. Other pharmaceutically active compounds that may be used can be found in Harrison's Principles of Internal Medicine, Thirteenth Edition, Eds. T.R. Harrison et al. McGraw-Hill N.Y., NY; and the Physicians Desk Reference 50th Edition 1997, Oradell New Jersey, Medical Economics Co., the complete contents of which are expressly incorporated herein by reference. The angiogenesis inhibitor compound and the pharmaceutically active compound may be administered to the subject in the same pharmaceutical composition or in different pharmaceutical compositions (at the same time or at different times).

Pharmaceutical Compositions of the Angiogenesis Inhibitor Compounds

The present invention also provides pharmaceutically acceptable formulations comprising one or more angiogenesis inhibitor compounds. Such pharmaceutically acceptable formulations typically include one or more angiogenesis inhibitor compounds as well as a pharmaceutically acceptable carrier(s) and/or excipient(s). As used herein, "pharmaceutically acceptable carrier" includes any and all solvents, dispersion media, coatings, antibacterial and anti fungal agents, isotonic and absorption delaying agents,

and the like that are physiologically compatible. The use of such media and agents for pharmaceutically active substances is well known in the art. Except insofar as any conventional media or agent is incompatible with the angiogenesis inhibitor compounds, use thereof in the pharmaceutical compositions is contemplated.

- 5 Supplementary pharmaceutically active compounds known to treat an angiogenic disease, *e.g.*, a chemotherapeutic agent such as Taxol, Paclitaxel, or Actinomycin D, or an antidiabetic agent such as Tolbutamide; or compounds that may potentiate the angiogenesis inhibitory activity of the angiogenesis inhibitor compound, such as heparin or a sulfated cyclodextrin, can also be incorporated into the compositions of the
- 10 invention. Suitable pharmaceutically active compounds that may be used can be found in Harrison's Principles of Internal Medicine (*supra*).

- A pharmaceutical composition of the invention is formulated to be compatible with its intended route of administration. Examples of routes of administration include parenteral, *e.g.*, intravenous, intradermal, subcutaneous, oral (*e.g.*, inhalation),
- 15 transdermal (topical), transmucosal, and rectal administration. Solutions or suspensions used for parenteral, intradermal, or subcutaneous application can include the following components: a sterile diluent such as water for injection, saline solution, fixed oils, polyethylene glycols, glycerine, propylene glycol or other synthetic solvents; antibacterial agents such as benzyl alcohol or methyl parabens; antioxidants such as
- 20 ascorbic acid or sodium bisulfite; chelating agents such as ethylenediaminetetraacetic acid; buffers such as acetates, citrates or phosphates and agents for the adjustment of tonicity such as sodium chloride or dextrose. pH can be adjusted with acids or bases, such as hydrochloric acid or sodium hydroxide. The parenteral preparation can be enclosed in ampoules, disposable syringes or multiple dose vials made of glass or
- 25 plastic.

- Pharmaceutical compositions suitable for injectable use include sterile aqueous solutions (where water soluble) or dispersions and sterile powders for the extemporaneous preparation of sterile injectable solutions or dispersion. For intravenous administration, suitable carriers include physiological saline, bacteriostatic
- 30 water, Cremophor EL™ (BASF, Parsippany, NJ) or phosphate buffered saline (PBS). In all cases, the pharmaceutical composition must be sterile and should be fluid to the extent that easy syringability exists. It must be stable under the conditions of manufacture and storage and must be preserved against the contaminating action of microorganisms such as bacteria and fungi. The carrier can be a solvent or dispersion
- 35 medium containing, for example, water, ethanol, polyol (for example, glycerol, propylene glycol, and liquid polyethylene glycol, and the like), and suitable mixtures thereof. The proper fluidity can be maintained, for example, by the use of a coating such as lecithin, by the maintenance of the required particle size in the case of dispersion and

by the use of surfactants. Prevention of the action of microorganisms can be achieved by various antibacterial and antifungal agents, for example, parabens, chlorobutanol, phenol, ascorbic acid, thimerosal, and the like. In many cases, it will be preferable to include isotonic agents, for example, sugars, polyalcohols such as manitol, sorbitol, sodium chloride in the composition. Prolonged absorption of the injectable compositions can be brought about by including in the composition an agent which delays absorption, for example, aluminum monostearate and gelatin.

Sterile injectable solutions can be prepared by incorporating the angiogenesis inhibitor compound in the required amount in an appropriate solvent with one or a combination of the ingredients enumerated above, as required, followed by filtered sterilization. Generally, dispersions are prepared by incorporating the angiogenesis inhibitor compound into a sterile vehicle which contains a basic dispersion medium and the required other ingredients from those enumerated above. In the case of sterile powders for the preparation of sterile injectable solutions, the preferred methods of preparation are vacuum drying and freeze-drying which yields a powder of the angiogenesis inhibitor compound plus any additional desired ingredient from a previously sterile-filtered solution thereof.

Oral compositions generally include an inert diluent or an edible carrier. They can be enclosed in gelatin capsules or compressed into tablets. For the purpose of oral therapeutic administration, the angiogenesis inhibitor compound can be incorporated with excipients and used in the form of tablets, troches, or capsules. Oral compositions can also include an enteric coating. Oral compositions can also be prepared using a fluid carrier for use as a mouthwash, wherein the angiogenesis inhibitor compound in the fluid carrier is applied orally and swished and expectorated or swallowed.

Pharmaceutically compatible binding agents, and/or adjuvant materials can be included as part of the composition. The tablets, pills, capsules, troches and the like can contain any of the following ingredients, or compounds of a similar nature: a binder such as microcrystalline cellulose, gum tragacanth or gelatin; an excipient such as starch or lactose, a disintegrating agent such as alginic acid, Primogel, or corn starch; a lubricant such as magnesium stearate or Sterotes; a glidant such as colloidal silicon dioxide; a sweetening agent such as sucrose or saccharin; or a flavoring agent such as peppermint, methyl salicylate, or orange flavoring.

For administration by inhalation, the angiogenesis inhibitor compounds are delivered in the form of an aerosol spray from pressured container or dispenser which contains a suitable propellant, *e.g.*, a gas such as carbon dioxide, or a nebulizer.

Systemic administration can also be by transmucosal or transdermal means. For transmucosal or transdermal administration, penetrants appropriate to the barrier to be permeated are used in the formulation. Such penetrants are generally known in the art,

and include, for example, for transmucosal administration, detergents, bile salts, and fusidic acid derivatives. Transmucosal administration can be accomplished through the use of nasal sprays or suppositories. For transdermal administration, the angiogenesis inhibitor compounds are formulated into ointments, salves, gels, or creams as generally known in the art.

The angiogenesis inhibitor compounds can also be prepared in the form of suppositories (*e.g.*, with conventional suppository bases such as cocoa butter and other glycerides) or retention enemas for rectal delivery.

In one embodiment, the angiogenesis inhibitor compounds are prepared with carriers that will protect the compound against rapid elimination from the body, such as a controlled release formulation, including implants and microencapsulated delivery systems. Biodegradable, biocompatible polymers can be used, such as ethylene vinyl acetate, polyanhydrides, polyglycolic acid, collagen, polyorthoesters, and polylactic acid. Methods for preparation of such formulations will be apparent to those skilled in the art. The materials can also be obtained commercially from Alza Corporation and Nova Pharmaceuticals, Inc. Liposomal suspensions can also be used as pharmaceutically acceptable carriers. These can be prepared according to methods known to those skilled in the art, for example, as described in U.S. Patent No. 4,522,811, U.S. Patent No. 5,455,044 and U.S. Patent No. 5,576,018, and U.S. Patent No. 4,883,666, the contents of all of which are incorporated herein by reference.

The angiogenesis inhibitor compounds of the invention can also be incorporated into pharmaceutical compositions which allow for the sustained delivery of the angiogenesis inhibitor compounds to a subject for a period of at least several weeks to a month or more. Such formulations are described in U.S. Patent 5,968,895, the contents of which are incorporated herein by reference.

It is especially advantageous to formulate oral or parenteral compositions in dosage unit form for ease of administration and uniformity of dosage. Dosage unit form as used herein refers to physically discrete units suited as unitary dosages for the subject to be treated; each unit containing a predetermined quantity of angiogenesis inhibitor compounds calculated to produce the desired therapeutic effect in association with the required pharmaceutical carrier. The specification for the dosage unit forms of the invention are dictated by and directly dependent on the unique characteristics of the angiogenesis inhibitor compound and the particular therapeutic effect to be achieved, and the limitations inherent in the art of compounding such angiogenesis inhibitor compounds for the treatment of individuals.

Toxicity and therapeutic efficacy of such angiogenesis inhibitor compounds can be determined by standard pharmaceutical procedures in cell cultures or experimental animals, *e.g.*, for determining the LD50 (the dose lethal to 50% of the population) and

the ED50 (the dose therapeutically effective in 50% of the population). The dose ratio between toxic and therapeutic effects is the therapeutic index and it can be expressed as the ratio LD50/ED50. Angiogenesis inhibitor compounds which exhibit large therapeutic indices are preferred. While angiogenesis inhibitor compounds that exhibit toxic side effects may be used, care should be taken to design a delivery system that targets such angiogenesis inhibitor compounds to the site of affected tissue in order to minimize potential damage to uninfected cells and, thereby, reduce side effects.

The data obtained from the cell culture assays and animal studies can be used in formulating a range of dosage for use in humans. The dosage of such angiogenesis inhibitor compounds lies preferably within a range of circulating concentrations that include the ED50 with little or no toxicity. The dosage may vary within this range depending upon the dosage form employed and the route of administration utilized. For any angiogenesis inhibitor compounds used in the methods of the invention, the therapeutically effective dose can be estimated initially from cell culture assays. A dose may be formulated in animal models to achieve a circulating plasma concentration range that includes the IC50 (*i.e.*, the concentration of the angiogenesis inhibitor compound which achieves a half-maximal inhibition of symptoms) as determined in cell culture. Such information can be used to more accurately determine useful doses in humans. Levels in plasma may be measured, for example, by high performance liquid chromatography.

Assays for Detecting the Activity of the Angiogenesis Inhibitor Compounds

The angiogenesis inhibitor compounds of the invention may be tested for their ability to modulate (*e.g.*, inhibit or stimulate) angiogenesis in a variety of well known assays, *e.g.*, the rat aortic ring angiogenesis inhibition assay (described herein in Example 27) or in a chorioallantoic membrane (CAM) assay.

The CAM assay may be performed essentially as described in Liekens S. et al. (1997) *Oncology Research* 9: 173-181, the contents of which are incorporated herein by reference. Briefly, fresh fertilized eggs are incubated for 3 days at 37°C. On the third day, the shell is cracked and the egg is placed into a tissue culture plate and incubated at 38°C. For the assay, the angiogenesis inhibitor compound to be tested is attached on a matrix of collagen on a nylon mesh. The mesh is then used to cover the chorioallantoic membrane and the eggs are incubated at 37°C. If angiogenesis occurs, new capillaries form and grow through the mesh within 24 hours. The ability of the angiogenesis inhibitor compound (at various concentrations) to modulate, *e.g.*, inhibit, angiogenesis, *e.g.*, FGF-induced angiogenesis, may then be determined.

The angiogenesis inhibitor compounds of the invention may also be tested for their ability to modulate (*e.g.*, inhibit or stimulate) human endothelial cell growth.

Human umbilical vein endothelial cells (HUVE) may be isolated by perfusion of an umbilical vein with a trypsin-containing medium. HUVE may then be cultured in GIT medium (Diago Eiyō Kagaku, Co., Japan) supplemented with 2.5% fetal bovine serum and 2.0 ng/ml of recombinant human basic fibroblast growth factor (rbFGF,

- 5 Biotechnology Research Laboratories, Takeda, Osaka, Japan) at 37°C under 5% CO₂ and 7% O₂. HUVE are then plated on 96-well microtiter plates (Nunc, 1-67008) at a cell density of 2×10^3 /100 µl of medium. The following day, 100 µl of medium containing rbFGF (2 ng/ml at the final concentration) and each angiogenesis inhibitor compound at various concentrations may be added to each well. The angiogenesis
- 10 inhibitor compounds are dissolved in dimethylsulfoxide (DMSO) and then diluted with culture medium so that the final DMSO concentration does not exceed 0.25%. After a 5-day culture, medium is removed, 100 µl of 1 mg/ml of MTT (3-(4,5-dimethyl-2-thiazolyl)-2,5-diphenyl-2 H-tetrazolium bromide) solution is added to the wells, and microtiters are kept at 37°C for 4 hours. Then, 100 µl of 10% sodium dodecyl sulfate
- 15 (SDS) solution is added to wells, and the microtiters are kept at 37°C for 5-6 hours. To determine the effects of the angiogenesis inhibitor compound on cell number, the optical density (590 µm) of each well is measured using an optical densitometer.

- The ability of the angiogenesis inhibitor compounds of the invention to modulate capillary endothelial cell migration *in vitro* may also be tested using the Boyden
- 20 chamber assay (as described in Falk *et al.* (1980) *J. Immunol. Meth.* 33:239-247, the contents of which are incorporated herein by reference). Briefly, bovine capillary endothelial cells are plated at 1.5×10^4 cells per well in serum-free DMEM (Dulbecco's Modified Eagle's Medium) on one side of nucleopore filters pre-coated with fibronectin (7.3 µg fibronectin/ml PBS). An angiogenesis inhibitor compound is
 - 25 dissolved in ethanol and diluted in DMEM so that the final concentration of ethanol does not exceed 0.01%. Cells are exposed to endothelial mitogen (Biomedical Technologies, Mass.) at 200 µg/ml and different concentrations of the angiogenesis inhibitor compound in serum-free DMEM for 4 hours at 37°C. At the end of this incubation, the number of cells that migrate through 8µ pores in the filters is determined by counting
 - 30 cells with an ocular grid at 100x in quadruplicate.

- The ability of the angiogenesis inhibitor compounds of the invention to modulate tumor growth may be tested *in vivo*. An animal model, *e.g.*, a C57BL/6N mouse with a mouse reticulum cell sarcoma (M 5076) intraperitoneally transplanted therein, may be used. The tumor cells in ascites can be collected by centrifugation, and suspended in
- 35 saline. The cell suspension (2×10^6 cells/100 µl/mouse) is inoculated into the right flanks of mice. Tumor-bearing mice are then subcutaneously treated with the angiogenesis inhibitor compound (at various concentrations suspended in 5% arabic gum solution containing 1% of ethanol) for 12 days beginning one day after the tumor inoculation.

The tumor growth may be determined by measuring tumor size in two directions with calipers at intervals of a few days.

Finally, the ability of the angiogenesis inhibitor compounds of the invention to modulate the activity of MetAP2 may be tested as follows. Recombinant human

- 5 MetAP2 may be expressed and purified from insect cells as described in Li and Chang, (1996) *Biochem. Biophys. Res. Commun.* 227:152-159. Various amounts of angiogenesis inhibitor compound is then added to buffer H (10 mM Hepes, pH 7.35, 100 mM KCl, 10% glycerol, and 0.1 M Co²⁺) containing 1nM purified recombinant human MetAP2 and incubated at 37°C for 30 minutes. To start the enzymatic reaction a peptide
10 containing a methionine residue, *e.g.*, Met-Gly-Met, is added to the reaction mixture (to a concentration of 1 mM). Released methionine is subsequently quantified at different time points (*e.g.*, at 0, 2, 3, and 5 minutes) using the method of Zou *et al.* (1995) *Mol. Gen Genetics* 246:247-253).

- 15 This invention is further illustrated by the following examples which should not be construed as limiting. The contents of all references, patents and published patent applications cited throughout this application, as well as the Figures and the Sequence Listing, are hereby incorporated by reference.

20

EXAMPLES

Synthetic methods

- 25 Compounds of the invention can be prepared using one or more of the following general methods.

- General Procedure A: To a mixture of carbonic acid-(3*R*, 4*S*, 5*S*, 6*R*)-5-methoxy-4-[(2*R*, 3*R*)-2-methyl-3-(3-methyl-but-2-enyl)-oxiranyl]-1-oxa-spiro[2.5]oct-6-yl ester 4-nitro-phenyl ester¹ (1, 0.47 mmol; Han, C. K.; Ahn, S. K.; Choi, N. S.; Hong, R. K.; Moon, S.
30 K.; Chun, H. S.; Lee, S. J.; Kim, J. W.; Hong, C. I.; Kim, D.; Yoon, J. H.; No, K. T. *Biorg. Med. Chem. Lett.* 2000, 10, 39-43) and amine (2.35 mmol) in EtOH (9 mL) was added dropwise, diisopropyl ethyl amine (2.35 mmol). After 3-18 hours, the ethanol was removed in vacuo and the crude material was dissolved into EtOAc (10 mL) and washed with H₂O (2 x 5 mL), and then brine (5 mL). The organic phase was dried over
35 Na₂SO₄ and the solvent removed in vacuo. Purification via flash chromatography (2-5% MeOH/CH₂Cl₂) afforded product.

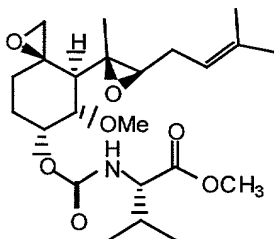
General Procedure B, Part I: A solution of (3*R*, 4*S*, 5*S*, 6*R*)-5-Methoxy-4-[(2*R*, 3*R*)-2-methyl-3-(3-methyl-but-2-enyl)-oxiranyl]-1-oxa-spiro[2.5]oct-6-yloxycarbonylamino)-acetic acid² (**2**, 0.11 mmol; U.S. Patent No. 6,017,954) in DMF (1 mL) was added to a 10 mL round bottomed flask containing swelled PS-DCC (0.28 mmol). In a separate vessel, the peptide (0.04 mmol) was dissolved into DMF (0.5 mL) and neutralized with NMM (0.04 mmol). After 1 hour, the solution of peptide was added to the pre-activated acid, and the reaction was continued for 5-18 hours. The resin was removed by filtration, washed with DMF (0.5 mL) and the solvent removed in vacuo. Purification via HPLC (CH₃CN/H₂O) afforded the product.

General Procedure B, Part II: A solution of the product in Part I (0.009 mmol) was dissolved into MeOH (1 mL) and was treated with Pd/C (2 mg), then subjected to a H₂ atmosphere (38 psi) for 24 hours. The mixture was then filtered through Celite, washed with MeOH (0.5 mL) and the solvent removed in vacuo. Purification via HPLC (CH₃CN/H₂O) afforded the product as a white solid.

General Procedure C: (1-Hydroxymethyl-methyl-propyl)-carbamic acid (3*R*, 4*S*, 5*S*, 6*R*)-5-methoxy-4-[(2*R*, 3*R*)-2-methyl-3-(3-methyl-but-2-enyl)-oxiranyl]-1-oxa-spiro[2.5]oct-6-yl ester (Example 7, 189 mg, 0.46 mmol), acid (0.46 mmol) and DMAP (0.69 mmol) were dissolved into anhydrous CH₂Cl₂ (5 mL) and treated with diisopropylcarbodiimide (0.46 mmol). After 7-18 hours, the solvent was removed in vacuo and purification via flash chromatography (MeOH/CH₂Cl₂) afforded the product.

Example 1

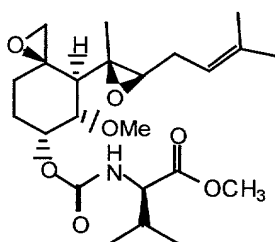
2-[(3*R*, 4*S*, 5*S*, 6*R*)-5-Methoxy-4-[(2*R*, 3*R*)-2-methyl-3-(3-methyl-but-2-enyl)-oxiranyl]-1-oxa-spiro[2.5]oct-6-yloxycarbonylamino]-3-methyl-butyric acid methyl ester



General procedure A was followed using **1** (31 mg, 0.07 mmol), L-valine methyl ester hydrochloride (58 mg, 0.35 mmol), and DIEA (60 μ L, 0.35 mmol) in EtOH (2 mL). Purification via flash chromatography (1% MeOH/ CH_2Cl_2) afforded the product as a clear oil (10 mg, 0.02 mmol, 33% yield); R_f = 0.60 (20% EtOAc/ CH_2Cl_2); LRMS (m/z) [M+1]⁺ 440.3 (calculated for $\text{C}_{23}\text{H}_{38}\text{NO}_7$, 440.3).

Example 2

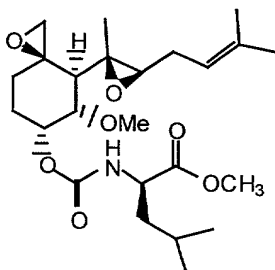
2-{(3*R*, 4*S*, 5*S*, 6*R*)-5-Methoxy-4-[(2*R*, 3*R*)-2-methyl-3-(3-methyl-but-2-enyl)-oxiranyl]-1-oxa-spiro[2.5]oct-6-yloxycarbonylamino}-3-methyl-butyric acid methyl ester



General procedure A was followed using **1** (41 mg, 0.09 mmol) and D-valine methyl ester hydrochloride (77 mg, 0.45 mmol), and DIEA (80 μ L, 0.45 mmol) in EtOH (2 mL). Purification via flash chromatography (1% MeOH/ CH_2Cl_2) afforded the product as a clear oil (18 mg, 0.04 mmol, 45% yield); R_f = 0.39 (20% EtOAc/ CH_2Cl_2); LRMS (m/z) [M+1]⁺ 440.3 (calculated for $\text{C}_{23}\text{H}_{38}\text{NO}_7$, 440.3).

Example 3

2-{(3*R*, 4*S*, 5*S*, 6*R*)-5-Methoxy-4-[(2*R*, 3*R*)-2-methyl-3-(3-methyl-but-2-enyl)-oxiranyl]-1-oxa-spiro[2.5]oct-6-yloxycarbonylamino}-4-methyl-pentanoic acid methyl ester



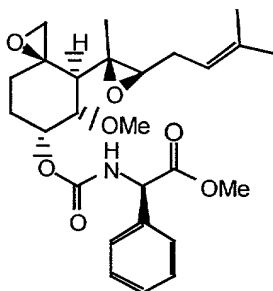
General procedure A was followed using **1** (23 mg, 0.05 mmol), D-leucine methyl ester hydrochloride (47 mg, 0.25 mmol), and DIEA (45 μ L, 0.25 mmol) in EtOH

(2 mL). Purification via flash chromatography (1% MeOH/CH₂Cl₂) afforded the product as a clear oil (19 mg, 0.04 mmol, 83% yield); *R*_f = 0.22 (15% EtOAc/CH₂Cl₂); LRMS (*m/z*) [M+1]⁺ 454.3 (calculated for C₂₄H₄₀NO₇, 454.3).

5

Example 4

{(3*R*, 4*S*, 5*S*, 6*R*)-5-Methoxy-4-[(2*R*, 3*R*)-2-methyl-3-(3-methyl-but-2-enyl)-oxiranyl]-1-oxa-spiro[2.5]oct-6-yloxycarbonylamino}-phenyl-acetic acid methyl ester



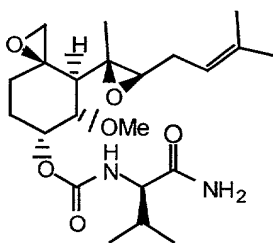
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General procedure A was followed using **1** (37 mg, 0.08 mmol), D-phenyl glycine methyl ester hydrochloride (83 mg, 0.40 mmol), and DIEA (72 μL, 0.40 mmol) in EtOH (2 mL). Purification via flash chromatography (1% MeOH/CH₂Cl₂) afforded the product as a clear oil (32 mg, 0.07 mmol, 82% yield); *R*_f = 0.41 (2% MeOH/CH₂Cl₂); LRMS (*m/z*) [M+1]⁺ 474.3 (calculated for C₂₆H₃₆NO₇, 474.3).

15

Example 5

(1-Carbamoyl-2-methyl-propyl)-carbamic acid-(3*R*, 4*S*, 5*S*, 6*R*)-5-methoxy-4-[(2*R*, 3*R*)-2-methyl-3-(3-methyl-but-2-enyl)-oxiranyl]-1-oxa-spiro[2.5]oct-6-yl ester

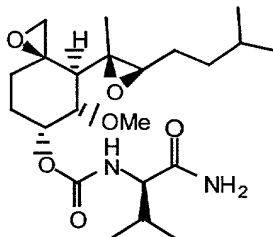


20

General procedure A was followed using **1** (55 mg, 0.12 mmol), D-valine amide hydrochloride (93 mg, 0.62 mmol), and DIEA (110 μL, 0.62 mmol) in EtOH (2 mL). Purification via flash chromatography (2% MeOH/CH₂Cl₂) afforded the product as a clear oil (42 mg, 0.10 mmol, 80% yield); *R*_f = 0.19 (2% MeOH/CH₂Cl₂); LRMS (*m/z*) [M+1]⁺ 425.5 (calculated for C₂₂H₃₇N₂O₆, 425.5).

Example 6

- 5 (1-Carbamoyl-2-methyl-propyl)-carbamic acid-(3*R*, 4*S*, 5*S*, 6*R*)-5-methoxy-4-[(2*R*, 3*R*)-2-methyl-3-(3-methyl-butyl)-oxiranyl]-1-oxa-spiro[2.5]oct-6-yl ester

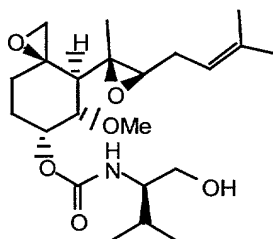


- 10 The compound in Example 4 (18 mg, 0.04 mmol) was dissolved into anhydrous MeOH (1.5 mL) and treated with Pd-C (2 mg) under a H₂ atmosphere. After 12 hours, the reaction was filtered through Celite and the solvent removed in vacuo to afford the product as a clear oil (18 mg, 0.04 mmol, 100% yield); *R*_f = 0.21 (2% MeOH/CH₂Cl₂); LRMS (*m/z*) [M+1]⁺ 427.5 (calculated for C₂₂H₃₉N₂O₆, 427.5).

15

Example 7

- (1-Hydroxymethyl-2-methyl-propyl)-carbamic acid-(3*R*, 4*S*, 5*S*, 6*R*)-5-methoxy-4-[(2*R*,3*R*)-2-methyl-3-(3-methyl-but-2-enyl)-oxiranyl]-1-oxa-spiro[2.5]oct-6-yl ester

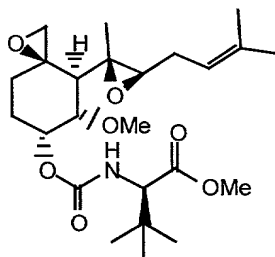


- 20 General procedure A was followed using **1** (290 mg, 0.65 mmol), D-valinol (337 mg, 3.25 mmol), and DIEA (560 μL, 3.25 mmol) in EtOH (5 mL). Purification via flash chromatography (2% MeOH/CH₂Cl₂) afforded the product as a clear oil (200 mg, 0.49 mmol, 75% yield); *R*_f = 0.26 (2% MeOH/CH₂Cl₂); LRMS (*m/z*) [M+1]⁺ 412.5 (calculated for C₂₂H₃₈NO₆, 412.5).

25

Example 8

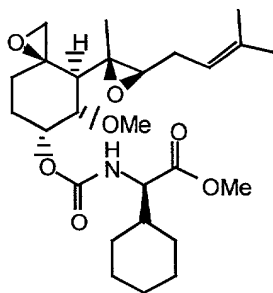
2- $\{(3R, 4S, 5S, 6R) \text{-} 5\text{-Methoxy-4-}[(2R, 3R)\text{-}2\text{-methyl-3-(3-methyl-but-2-enyl)-oxiranyl}]\text{-1-oxa-spiro[2.5]oct-6-yloxy-carbonylamino}\}$ -3,3-dimethyl-but-2-enoic acid methyl ester



General procedure A was followed using **1** (65 mg, 0.15 mmol), D-tBu glycine methyl ester hydrochloride (132 mg, 0.73 mmol), and DIEA (127 μ L, 0.73 mmol) in EtOH (8 mL). Purification via flash chromatography (10% EtOAc/ CH_2Cl_2) afforded the product as a clear oil (10 mg, 0.02 mmol, 15% yield); R_f = 0.22 (10% EtOAc/ CH_2Cl_2); LRMS (m/z) $[\text{M}+1]^+$ 454.5 (calculated for $\text{C}_{24}\text{H}_{40}\text{NO}_7$, 454.5).

Example 9

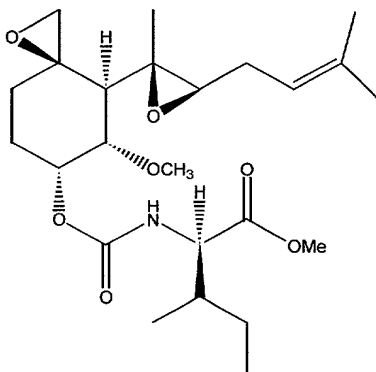
Cyclohexyl-2- $\{(3R, 4S, 5S, 6R) \text{-} 5\text{-Methoxy-4-}[(2R, 3R)\text{-}2\text{-methyl-3-(3-methyl-but-2-enyl)-oxiranyl}]\text{-1-oxa-spiro[2.5]oct-6-yloxy-carbonylamino}\}$ -acetic acid methyl ester



General procedure A was followed using **1** (65 mg, 0.15 mmol), D-cyclohexyl glycine methyl ester hydrochloride (207 mg, 0.73 mmol), and DIEA (127 μ L, 0.73 mmol) in EtOH (7 mL). Purification via flash chromatography (10% EtOAc/ CH_2Cl_2) afforded the product as a clear oil (20 mg, 0.04 mmol, 28% yield); R_f = 0.22 (10% EtOAc/ CH_2Cl_2); LRMS (m/z) $[\text{M}+1]^+$ 480.3 (calculated for $\text{C}_{26}\text{H}_{42}\text{NO}_7$, 480.3).

Example 10

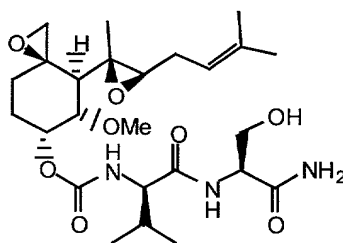
2-[(3*R*, 4*S*, 5*S*, 6*R*)-5-Methoxy-4-[(2*R*, 3*R*)-2-methyl-3-(3-methyl-but-2-enyl)-oxiranyl]-1-oxa-spiro[2.5]oct-6-yloxycarbonylamino]-3-methyl-pentanoic acid methyl ester



5

- General procedure A was followed using **1** (65 mg, 0.15 mmol), D-isoleucine methyl ester hydrochloride (132 mg, 0.73 mmol), and DIEA (127 μ L, 0.73 mmol) in EtOH (7 mL). Purification via flash chromatography (10% EtOAc/ CH_2Cl_2) afforded the product as a clear oil (20 mg, 0.04 mmol, 30% yield); R_f = 0.20 (10% EtOAc/ CH_2Cl_2); LRMS (m/z) $[\text{M}+1]^+$ 454.5 (calculated for $\text{C}_{24}\text{H}_{40}\text{NO}_7$, 454.5).

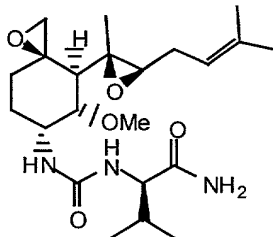
- Example 11**
- 15 [1-(1-Carbamoyl-2-hydroxy-ethylcarbamoyl)-2-methyl-propyl]-carbamic acid-(3*R*, 4*S*, 5*S*, 6*R*)-5-methoxy-4-[(2*R*, 3*R*)-2-methyl-3-(3-methyl-but-2-enyl)-oxiranyl-1-oxa-spiro[2.5]oct-6-yl ester



- General procedure A was followed using **1** (74 mg, 0.17 mmol), H-D-vS- $\text{NH}_2 \cdot \text{TFA}$ (262 mg, 0.83 mmol), and DIEA (140 μ L, 0.83 mmol) in EtOH (5 mL). Purification via HPLC (60% $\text{CH}_3\text{CN}/\text{H}_2\text{O}$) afforded the as a white solid (34 mg, 0.07 mmol, 40% yield); R_f = 0.21 (5% MeOH/ CH_2Cl_2); LRMS (m/z) $[\text{M}+1]^+$ 512.5 (calculated for $\text{C}_{25}\text{H}_{42}\text{N}_3\text{O}_8$, 512.3).

Example 12

2-(3-((3*R*, 4*S*, 5*S*, 6*R*)-5-Methoxy-4-[(2*R*, 3*R*)-2-methyl-3-(3-methyl-but-2-enyl)-oxiranyl]-1-oxa-spiro[2.5]oct-6-yl}-ureido)-3-methyl-butamide

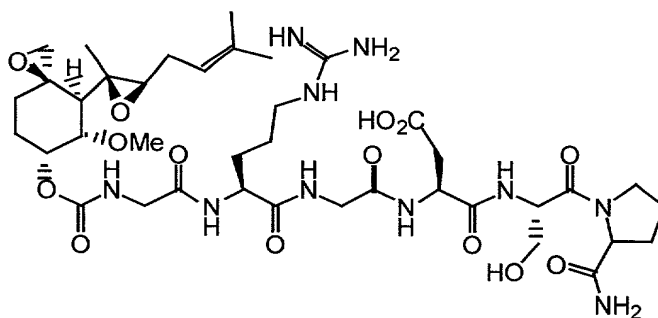


- 5 (3*R*, 4*S*, 5*S*, 6*R*)-5-Methoxy-4-[(2*R*, 3*R*)-2-methyl-3-(3-methyl-but-2-enyl)-oxiranyl]-1-oxa-spiro[2.5]oct-6-ylamine (**3**; PCT Publication No. WO 99/59987) was prepared according to the published procedure. To a solution of crude **3** (29 mg, 0.1 mmol), DIEA (21 mL, 0.1 mmol) and DMAP (2 mg) in CH₂Cl₂ (1.5 mL) cooled to 0 °C was added p-NO₂ phenyl chloroformate (25 mg, 0.12 mmol). After 45 minutes, the
10 reaction was warmed to room temperature and a solution of H-D-val-NH₂•HCl (40 mg, 0.15 mmol) in EtOH (1 mL) and DIEA (35 µL, 0.2 mmol) was added.

- The reaction was continued for 1 hour, then was concentrated in vacuo, taken up into EtOAc (15 mL), and washed with dilute HCl_{aq} (2 x 15 mL), H₂O (2 x 15 mL) and brine (15 mL). Purification via flash chromatography (5% MeOH/CH₂Cl₂) afforded the
15 product as a white solid (6 mg, 0.014 mmol, 13% yield from **3**); R_f = 0.12 (5% MeOH/CH₂Cl₂); LRMS (*m/z*) [M+1]⁺ 424.4 (calculated for C₂₂H₃₈N₃O₅, 424.4).

Example 13

- 20 N-Carbamoyl (ID#31) 3*R*, 4*S*, 5*S*, 6*R* 5-methoxy-4-[(2*R*,3*R*)2-methyl-3-(3-methyl-butyl)-oxiranyl]-1-oxa-spiro[2.5]oct-6-yl ester



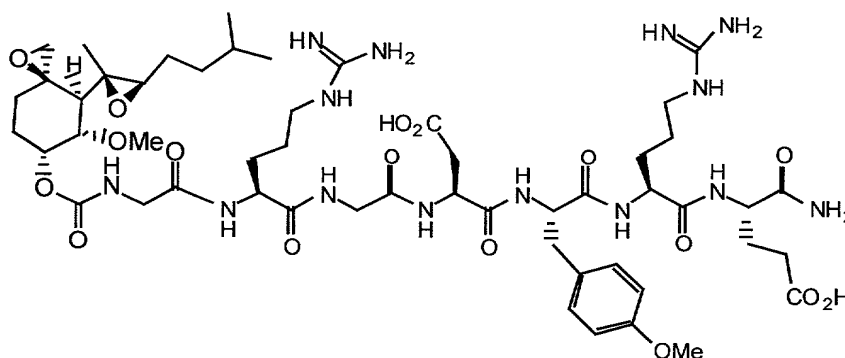
General Procedure B, Part I was followed using **2** (41 mg, 0.11 mmol), PS-DCC (256 mg, 0.28 mmol) in DMF (1 mL) and H-RGD(Bn)S(OBn)P-NH₂•2TFA (37 mg, 0.04 mmol), NMM (4 µL, 0.04 mmol) in DMF (0.5 mL). Purification via HPLC

(70%CH₃CN/H₂O/0.075% TFA) afforded the product as a white flocculent solid (9.3 mg, 0.009 mmol, 17%yield); LRMS (*m/z*) [M+1]⁺ 1075.4 (calculated for C₅₃H₇₅N₁₀O₁₄, 1075.5).

General Procedure, Part II was followed using the product in Part I (9.3 mg, 0.009 mmol) and Pd/C (2 mg) in MeOH (1 mL), and a H₂ atmosphere (38 psi) for 24 hours. Purification via HPLC (55%CH₃CN/H₂O/0.075% TFA) afforded the product as a white solid (5 mg, 0.006 mmol, 65% yield); LRMS (*m/z*) [M+1]⁺ 897.3 (calculated for C₃₉H₆₅N₁₀O₁₄, 897.5).

Example 14

N-Carbamoyl (ID#30) 3*R*, 4*S*, 5*S*, 6*R*) 5-methoxy-4-[(2*R*,3*R*)2-methyl-3-(3-methyl-butyl)-oxiranyl]-1-oxa-spiro[2.5]oct-6-yl ester



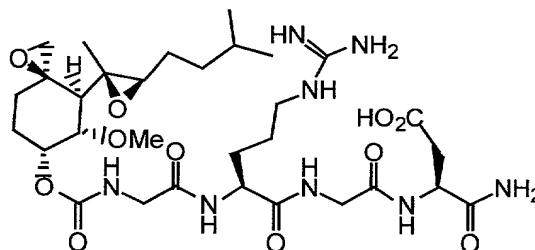
General Procedure, Part I was followed using **2** (38 mg, 0.10 mmol) and PS-DCC (238 mg, 0.25 mmol) in DMF (1 mL), H-RGD(Bn)Y(OMe)RE(Bn)-NH₂•3TFA (35 mg, 0.03 mmol) and NMM (3 μL, 0.03 mmol) in DMF (0.5 mL). Purification via HPLC (70%CH₃CN/H₂O/0.075% TFA) afforded the product as a white flocculent solid (4.0 mg, 0.002 mmol, 8%yield); LRMS (*m/z*) [M+2/2]⁺ 677.6 (calculated for C₆₆H₉₂N₁₄O₁₇, 677.8).

General Procedure, Part II was followed using the product in Part I (3.0 mg, 0.002 mmol) and Pd/C (2 mg) in MeOH (1 mL), under a H₂ atmosphere (38 psi) for 24 hours. Purification via HPLC (55%CH₃CN/H₂O/0.075% TFA) afforded the product as a white solid (3.3 mg, 0.0027 mmol, 94% yield); LRMS (*m/z*) [M+2/2]⁺ 588.5 (calculated for C₅₂H₈₂N₁₄O₁₇, 588.7).

Example 15

N-Carbamoyl (ID#32) 3*R*, 4*S*, 5*S*, 6*R*) 5-methoxy-4-[(2*R*,3*R*)2-methyl-3-(3-methyl-butyl)-oxiranyl]-1-oxa-spiro[2.5]oct-6-yl ester

5



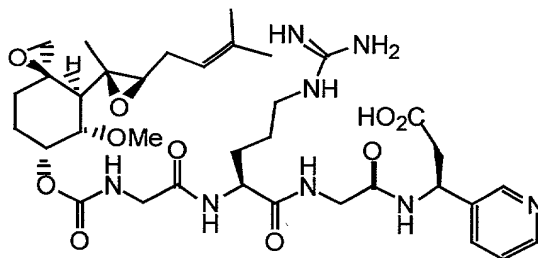
General Procedure B, Part I was followed using **2** (38 mg, 0.10 mmol), PS-DCC (238 mg, 0.25 mmol), and HOBT (29 mg, 0.25 mmol) in DMF (1 mL), and H-
 10 RGDBnNH₂•TFA (29 mg, 0.04 mmol) and NMM (4.8 μL, 0.04 mmol) in DMF (0.5 mL). Purification via HPLC (60%CH₃CN/H₂O/0.075% TFA) afforded the product as a white solid (35 mg, 0.04 mmol, 44% yield); LRMS (*m/z*) 801.2 (calculated for C₃₈H₅₇N₈O₁₁, 801.4).

General Procedure, Part II was followed using the product in Part I (35 mg, 0.04
 15 mmol) and Pd/C (2 mg) in MeOH (1 mL), under a H₂ atmosphere (38 psi) for 24 hours. Purification via HPLC (50%CH₃CN/H₂O/0.075% TFA) afforded the product as a white solid (22 mg, 0.03 mmol, 71% yield); LRMS (*m/z*) 713.2 (calculated for C₃₁H₅₃N₈O₁₁, 713.4).

20

Example 16

N-Carbamoyl (ID#40) (3*R*, 4*S*, 5*S*, 6*R*) 5-methoxy-4-[(2*R*,3*R*)2-methyl-3-(3-methyl-but-2-enyl)-oxiranyl]-1-oxa-spiro[2.5]oct-6-yl ester



25

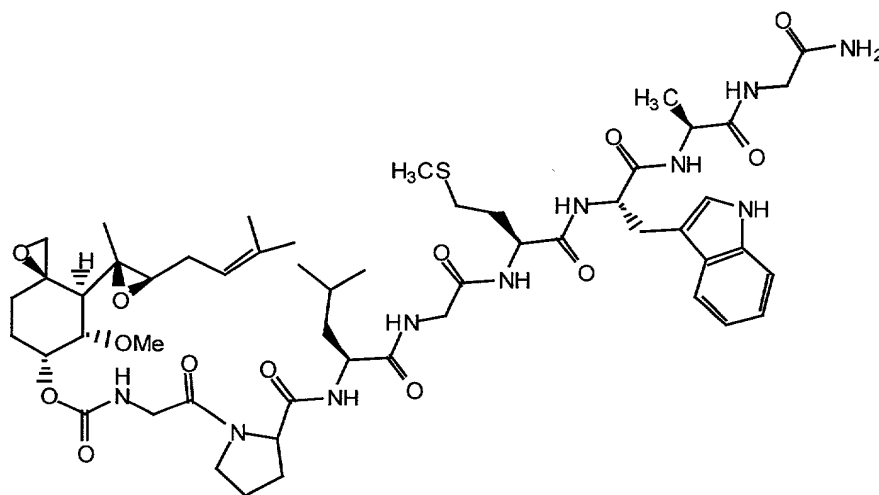
General Procedure B, Part I was followed using **2** (65 mg, 0.17 mmol), PS-DCC (405 mg, 0.43 mmol), and HOBT (34 mg, 0.26 mmol) in DMF (1 mL), and H-RG(pyridyl)D-OMe (43 mg, 0.06 mmol) and NMM (7 μL, 0.06 mmol) in DMF (0.5

mL). Purification via HPLC (50% CH₃CN/H₂O/0.075% TFA) afforded the product as a white solid (15 mg, 0.02 mmol, 34% yield); LRMS (*m/z*) 773.2 (calculated for C₃₄H₅₅N₄O₁₀, 773.4).

The product of Part I (11 mg, 0.01 mmol) was dissolved into THF:MeOH:H₂O (2:1:1, 500 μL) and treated with LiOH·H₂O (1.2 mg, 0.02 mmol) for 2 hours. The crude material was diluted with EtOAc (5 mL) and acidified with dilute HCl (10 mL). The aqueous phase was washed with additional EtOAc (2 x 5 mL), the combined organic extracts dried over Na₂SO₄ and the solvent removed in vacuo. Purification via HPLC (30% CH₃CN/H₂O/0.075% TFA) afforded the product as a white solid (2 mg, 0.003 mmol, 19% yield). LRMS (*m/z*) 745.3 (calculated for C₃₃H₅₃N₄O₁₀, 745.4).

Example 17

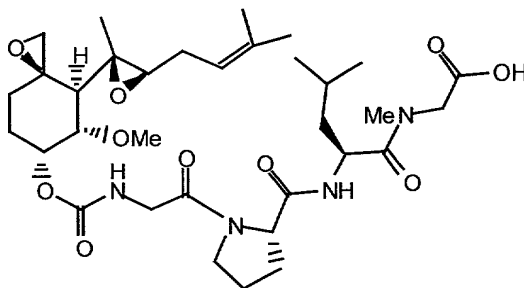
N-Carbamoyl (ID#39) (3*R*, 4*S*, 5*S*, 6*R*) 5-methoxy-4-[(2*R*,3*R*)2-methyl-3-(3-methyl-but-2-enyl)-oxiranyl]-1-oxa-spiro[2.5]oct-6-yl ester



General procedure B, Part I was followed using **2** (25 mg, 0.07 mmol) and PS-DCC (155 mg, 0.16 mmol) in DMF (1 mL), and H-PLGMWAG-NH₂ (20 mg, 0.03 mmol) and NMM (3 μL, 0.03 mmol) in DMF (0.5 mL). Purification via HPLC (70%CH₃CN/H₂O/0.075% TFA) afforded the product as a white solid (1.4 mg, 0.001 mmol, 5% yield); LRMS (*m/z*) [M+1]⁺ 1095.6 (calculated for C₅₃H₇₉N₁₀O₁₃S, 1095.6).

Example 18

N-Carbamoyl (ID#26) (3*R*, 4*S*, 5*S*, 6*R*) 5-methoxy-4-[(2*R*,3*R*)2-methyl-3-(3-methyl-but-2-enyl)-oxiranyl]-1-oxa-spiro[2.5]oct-6-yl ester



5

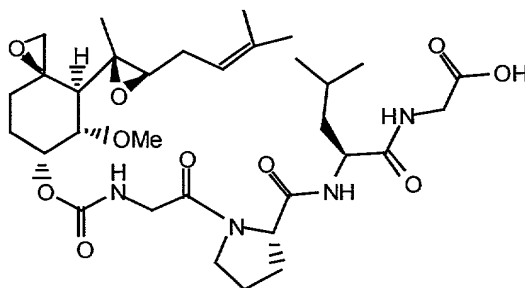
General Procedure B, Part I was followed using **2** (69 mg, 0.18 mmol), PS-DCC (429 mg, 0.45 mmol) and HOBT (21 mg, 0.18 mmol) in DMF (1 mL), and H-PL(N-Me)G-OMe (31 mg, 0.07 mmol) and NMM (8 μ L, 0.07 mmol) in DMF (0.5 mL). Purification via HPLC (70%CH₃CN/H₂O/0.075% TFA) afforded the product as a white solid (27 mg, 0.04 mmol, 59%yield); LRMS (*m/z*) [M+1]⁺ 679.4 (calculated for C₃₄H₅₅N₄O₁₀, 679.4).

The product of Part I (27 mg, 0.04 mmol) was dissolved into THF:MeOH:H₂O (2:1:1, 1.5 mL) and treated with LiOH•H₂O (4 mg, 0.10 mmol) for 1 hour. The solution was acidified to pH 3 using 0.1 N HCl, and the MeOH and THF removed in vacuo. Purification via HPLC (60% CH₃CN/H₂O/0.075% TFA) afforded the product as a white solid (6 mg, 0.01 mmol, 23% yield). LRMS (*m/z*) 665.4 (calculated for C₃₃H₅₃N₄O₁₀, 665.4).

20

Example 19

N-Carbamoyl (ID#27) (3*R*, 4*S*, 5*S*, 6*R*) 5-methoxy-4-[(2*R*,3*R*)2-methyl-3-(3-methyl-but-2-enyl)-oxiranyl]-1-oxa-spiro[2.5]oct-6-yl ester

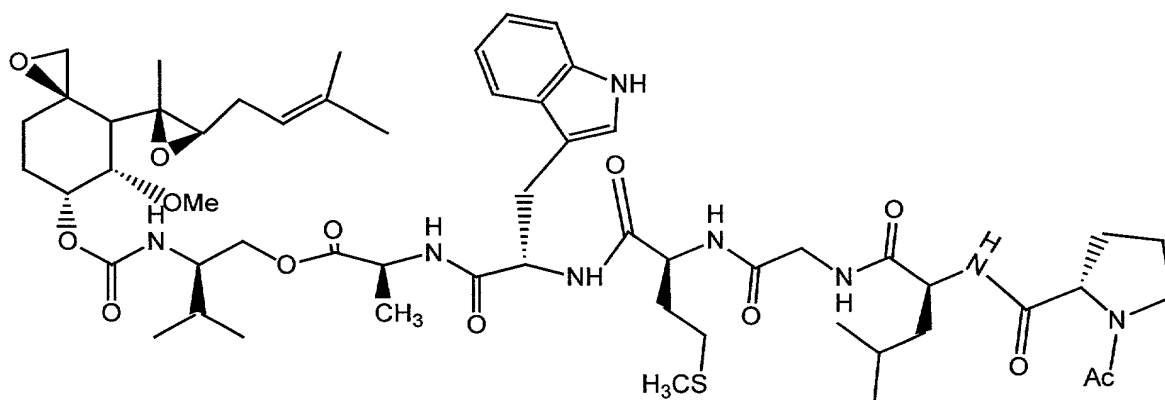


General Procedure B, Part I was followed using **2** (44 mg, 0.12 mmol), PS-DCC (276 mg, 0.29 mmol) and HOBt (27 mg, 0.23 mmol) in DMF (1 mL), and H-PLG-OMe (20 mg, 0.05 mmol) and NMM (5 μ L, 0.05 mmol) in DMF (0.5 mL). Purification via HPLC (90%CH₃CN/H₂O/0.075% TFA) afforded the product as a white solid (13 mg, 0.02 mmol, 43%yield); LRMS (*m/z*) [M+1]⁺ 664.4 (calculated for C₃₄H₅₂N₄O₁₀, 664.4).

The product in Part I (27 mg, 0.04 mmol) was dissolved into THF:MeOH:H₂O (2:1:1, 790 μ L) and treated with LiOH•H₂O (1.2 mg, 0.03 mmol) for 2 hours. The solution was acidified to pH 3 using 0.1 N HCl, and the MeOH and THF removed in vacuo. Purification via HPLC (90% CH₃CN/H₂O/0.075% TFA) afforded the product as a white solid (1.8 mg, 0.003 mmol, 15% yield). LRMS (*m/z*) 650.4 (calculated for C₃₂H₅₀N₄O₁₀, 650.4).

Example 20

(ID#24)-(2*R*)-{(3*R*, 4*S*, 5*S*, 6*R*) 5-methoxy-4-[(2*R*,3*R*)2-methyl-3-(3-methyl-but-2-enyl)-oxiranyl]-1-oxa-spiro[2.5]oct-6-yloxycarbonyl} amino-3-methyl-butanol) ester

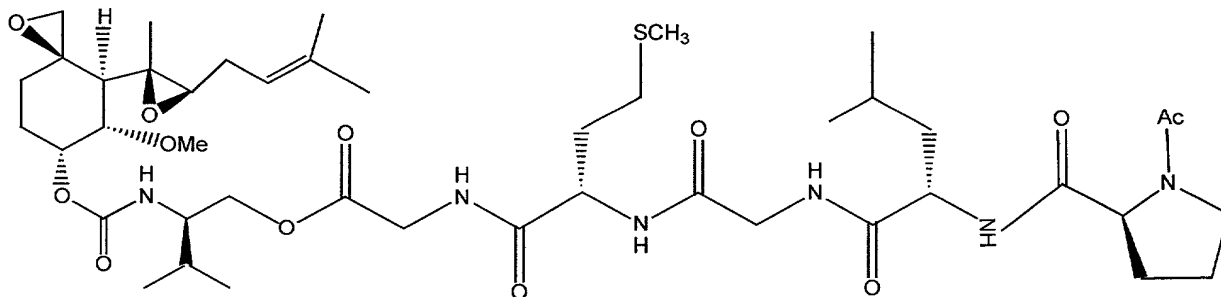


General Procedure C was followed using the compound in Example 7 (189 mg, 0.46 mmol), Ac-PLGMWA-OH (329 mg, 0.46 mmol), DMAP (84 mg, 0.69 mmol) and DIC (72 μ L, 0.46 mmol) in CH₂Cl₂ (5 mL). After 18 hours, the solvent was removed in vacuo and purification via flash chromatography (2% MeOH/CH₂Cl₂) afforded the product as a white solid (357 mg, 0.32 mmol, 70% yield); R_f = 0.18 (5% MeOH/CH₂Cl₂); LRMS (*m/z*) [M+1]⁺ 1110.3 (calculated for C₅₆H₈₅N₈O₁₃S, 1110.3).

Example 21

(ID#36)-(2*R*-(3*R*, 4*S*, 5*S*, 6*R*) 5-methoxy-4-[(2*R*,3*R*)2-methyl-3-(3-methyl-but-2-enyl)-oxiranyl]-1-oxa-spiro[2.5]oct-6-yloxycarbonyl} amino-3-methyl-butanol) ester

5

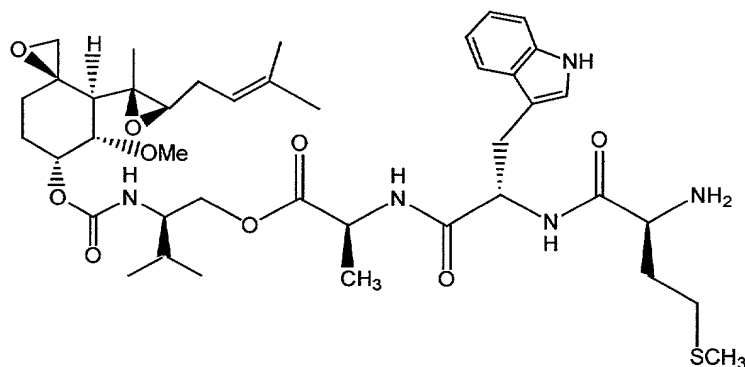


General Procedure C was followed using the compound in Example 7 (61 mg, 0.15 mmol), Ac-PLGMG-OH (92 mg, 0.18 mmol), DMAP (22 mg, 0.18 mmol) and DIC (28 μ L, 0.18 mmol) in CH_2Cl_2 (2 mL). After 7 hours, the solvent was removed in vacuo and purification via flash chromatography (3% MeOH/ CH_2Cl_2) afforded the product as a white solid (61 mg, 0. mmol, 45% yield); R_f = 0.20 (5% MeOH/ CH_2Cl_2); LRMS (m/z) $[M+1]^+$ 909.7 (calculated for $\text{C}_{44}\text{H}_{73}\text{N}_6\text{O}_{12}\text{S}$, 909.5).

15

Example 22

(ID#37)-(2*R*-(3*R*, 4*S*, 5*S*, 6*R*) 5-methoxy-4-[(2*R*,3*R*)2-methyl-3-(3-methyl-but-2-enyl)-oxiranyl]-1-oxa-spiro[2.5]oct-6-yloxycarbonyl} amino-3-methyl-butanol) ester



20

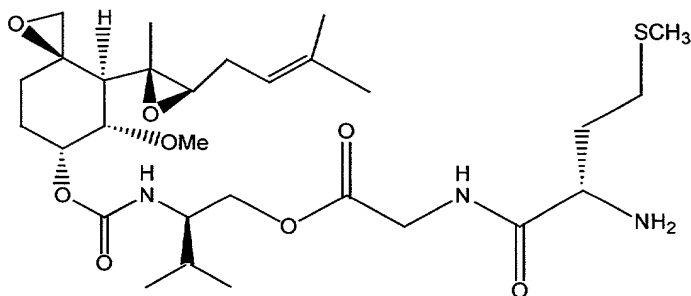
General Procedure C was followed using the compound in Example 7 (79 mg, 0.19 mmol), Fmoc-MWA-OH (121 mg, 0.19 mmol) and DMAP (4 mg, 0.03 mmol) and

DIC (30 μ L, 0.19 mmol) in CH_2Cl_2 (2 mL). After 11 hours, the solvent was removed in vacuo and purification via flash chromatography (2% MeOH/ CH_2Cl_2) afforded the product as a white solid (128 mg, 0.12 mmol, 65% yield); LRMS (m/z) $[\text{M}+1]^+$ 1022.9 (calculated for $\text{C}_{44}\text{H}_{73}\text{N}_6\text{O}_{12}\text{S}$, 1022.5).

- 5 The product from General Procedure C (above) (54 mg, 0.05 mmol) was dissolved into anhydrous CH_2Cl_2 (3 mL) cooled to 0 $^\circ\text{C}$, then treated with a gentle stream of $\text{NH}_{3(\text{g})}$ for 15 minutes. The reaction was sealed and continued at 0 $^\circ\text{C}$ for 36 hours. The solvent was removed in vacuo, and the crude residue acidified with $\text{CH}_3\text{CN}/\text{H}_2\text{O}$ (0.075% TFA) (5 mL). Purification via HPLC (70% $\text{CH}_3\text{CN}/\text{H}_2\text{O}$ /0.075% TFA) afforded the product as a white solid (2 mg, 0.003 mmol, 5% yield); LRMS (m/z) $[\text{M}+1]^+$ 800.6 (calculated for $\text{C}_{41}\text{H}_{62}\text{N}_5\text{O}_9\text{S}$, 800.5).

Example 23

- (ID#38)-(2*R*-(3*R*, 4*S*, 5*S*, 6*R*) 5-methoxy-4-[(2*R*,3*R*)2-methyl-3-(3-methyl-but-2-enyl)-oxiran-1-yl]-1-oxa-spiro[2.5]oct-6-yl)oxycarbonyl} amino-3-methyl-butanol) ester

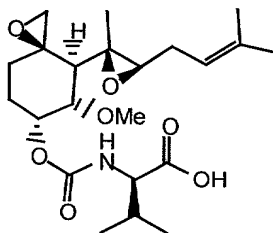


- General Procedure C was followed using the compound in Example 7 (76 mg, 0.18 mmol), Fmoc-MG-OH (79 mg, 0.18 mmol) and DMAP (4 mg, 0.03 mmol) and VDIC (29 μ L, 0.18 mmol) in CH_2Cl_2 (2 mL). After 10 hours, the solvent was removed in vacuo and purification via flash chromatography (2% MeOH/ CH_2Cl_2) afforded the product as a white solid (128 mg, 0.12 mmol, 65% yield); LRMS (m/z) $[\text{M}+1]^+$ 822.6 (calculated for $\text{C}_{44}\text{H}_{60}\text{N}_3\text{O}_{10}\text{S}$, 822.5).

- 25 The product from General Procedure C (above) (42 mg, 0.05 mmol) was dissolved into anhydrous CH_2Cl_2 (3 mL) cooled to 0 $^\circ\text{C}$, then treated with a gentle stream of $\text{NH}_{3(\text{g})}$ for 15 minutes. The reaction was sealed and continued at 0 $^\circ\text{C}$ for 36 hours. The solvent was removed in vacuo, and the crude residue acidified with $\text{CH}_3\text{CN}/\text{H}_2\text{O}$ (0.075% TFA) (5 mL). Purification via HPLC (70% $\text{CH}_3\text{CN}/\text{H}_2\text{O}$ /0.075% TFA) afforded the product as a white solid (2 mg, 0.003 mmol, 5% yield); LRMS (m/z) $[\text{M}+1]^+$ 600.4 (calculated for $\text{C}_{29}\text{H}_{50}\text{N}_3\text{O}_8\text{S}$, 600.4).

Example 24

2-{(3*R*, 4*S*, 5*S*, 6*R*)-5-Methoxy-4-[(2*R*, 3*R*)-2-methyl-3-(3-methyl-but-2-enyl)-oxiranyl]-1-oxa-spiro[2.5]oct-6-yloxy-carbonylamino}-3-methyl-but-2-enoic acid



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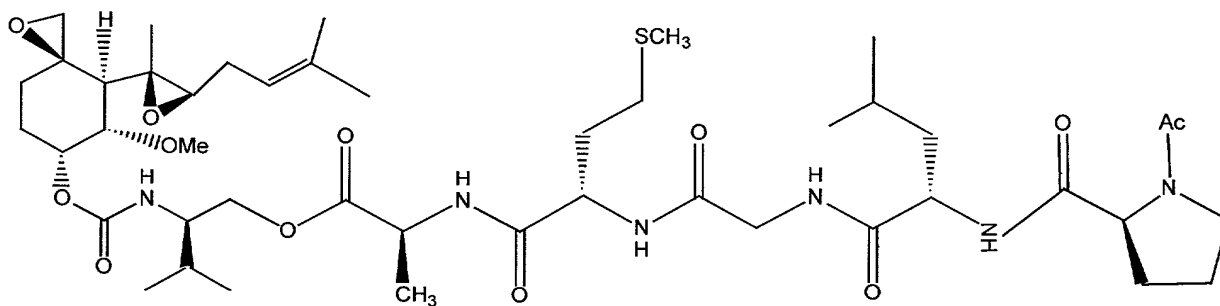
The compound in Example 2 (9 mg, 0.02 mmol) was dissolved into THF:MeOH:H₂O (1 mL) and treated with LiOH·H₂O (2 mg, 0.05 mmol). After 2 hours, the reaction was partitioned between EtOAc (5 mL) and dilute HCl (5 mL). The organic phase was dried over Na₂SO₄ and the solvent removed in vacuo. Purification via HPLC (85% CH₃CN/H₂O/0.075% TFA) afforded the product as a white solid (0.58 mg, 0.001 mmol, 6% yield); LRMS (*m/z*) [M+1]⁺ 426.4 (calculated for C₂₂H₃₆NO₇, 426.5).

10

Example 25

(ID#34)-(2*R*)-{(3*R*, 4*S*, 5*S*, 6*R*) 5-methoxy-4-[(2*R*,3*R*)2-methyl-3-(3-methyl-but-2-enyl)-oxiranyl]-1-oxa-spiro[2.5]oct-6-yloxy-carbonyl} amino-3-methyl-butanol ester

15



General Procedure C was followed using the compound in Example 7 (41 mg, 0.10 mmol), Ac-PLGMG-OH (63 mg, 0.12 mmol), DMAP (15 mg, 0.12 mmol) and DIC (19 μL, 0.12 mmol) in CH₂Cl₂ (2 mL). After 7 hours, the solvent was removed in vacuo and purification via flash chromatography (3% MeOH/CH₂Cl₂) afforded the product as a white solid (43 mg, 0.05 mmol, 47% yield); R_f = 0.21 (5% MeOH/CH₂Cl₂); LRMS (*m/z*) [M+1]⁺ 923.7 (calculated for C₄₅H₇₅N₆O₁₂S, 923.5).

20

Example 26

The angiogenesis inhibitor compounds of the invention were tested for their ability to modulate human endothelial cell growth and for their ability to modulate the activity of MetAP2. The MetAP2 enzyme assay was performed essentially as described in Turk, B. *et al.* (1999) *Chem. & Bio.* 6: 823-833, the entire contents of which are incorporated herein by reference. The bovine aortic endothelial cell growth assay (Baec assay) was performed essentially as described in Turk, B. *et al.* (*supra*), the entire contents of which are incorporated herein by reference.

For the human endothelial cell growth assay, human umbilical vein endothelial cells (HUVEC) were maintained in Clonetics endothelial growth medium (EGM) in a 37 °C humidified incubator. Cells were detached with trypsin and pelleted by centrifugation at 300 x g for 5 minutes at room temperature. HUVEC were added to 96-well plates at 5,000 cells/well. After incubating for 6 hours, the medium was replaced with 0.2 ml fresh EGM supplemented with 0.5 nM bFGF and the desired concentration of test angiogenesis inhibitor compound. Test angiogenesis inhibitor compounds were initially dissolved in ethanol at stock concentrations of either 10 mM or 0.1 mM, and subsequently diluted in EGM to obtain concentrations from 1 pM to 10 μ M. After 48 hours at 37 °C, the medium was replaced with fresh bFGF-supplemented EGM and test angiogenesis inhibitor compound. Following incubation for an additional 48 hours at 37 °C MTT (3-[4,5-dimethylthiazol-2-yl]-2,5-diphenyl-tetrazolium bromide) was added to 1 mg/ml. After 2-4 hours at 37 °C the medium was replaced with 0.1 ml/well isopropanol. The plates were placed on a shaker for 15 minutes at room temperature and analyzed in a Labsystems Multiskan plate spectrophotometer at an optical density of 570 nm.

The results of the assays, set forth below in Tables I-III, demonstrate that the angiogenesis inhibitor compounds of the invention have excellent MetAP2 inhibitory activity and are able to inhibit endothelial cell growth at the picomolar range.

Table I. MetAP2 Assay

Example	IC ₅₀ (nM)
1	4.7
2	2
3	5.5
4	2.7
13	2.9
14	4000
17	16.7

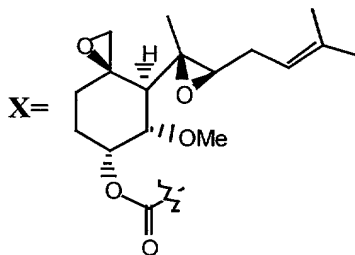
Table II. Huvec Assay

Example	IC ₅₀ (pM)
1	18
2	40
3	38
4	36
5	93
13	(>10 μ M)
14	(>10 μ M)
15	(>10 μ M)
17	(95 nM)
18	(>100 nM)
19	(>100 nM)
24	5444

5 **Table III.** Baec Assay

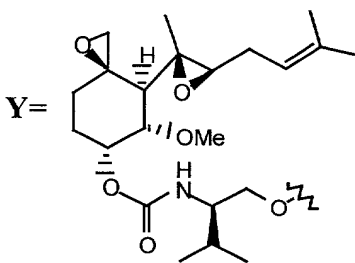
Example	IC ₅₀ (pM)
1	17
2	48
3	118
4	35
5	46
6	220
7	128
8	313
9	165
10	179
11	(>100 nM)
16	(>100 nM)
19	(>100 nM)
22	326
23	207

The identity of the angiogenesis inhibitor compounds used in each of the experiments is shown in Tables IV and V below.

Table IV.

5

Example	ID#	Sequence
13	31	X -GlyArgGlyAspSerPro-NH ₂
14	30	X -GlyArgGlyAspTyr(OMe)Arg Glu-NH ₂
15	32	X -GlyArgGlyAsp-NH ₂
16	40	X -GlyArg{3-amino-(3-pyridyl)}-propionic acid
17	39	X -GlyProLeuGlyMetTrpAlaGly-NH ₂
18	26	X -GlyProLeuSar-OH
19	27	X -GlyProLeuGly-OH

Table V.

Example	ID#	Sequence
20	24	Ac-ProLeuGly-MetTrpAla- Y
21	36	Ac-ProLeuGlyMetGly- Y
22	37	H-MetTrpAla- Y
23	38	H-MetGly- Y
25	34	Ac-ProLeuGlyMetAla- Y

10

Example 27

5 The angiogenesis inhibitor compounds of the invention were also tested for their ability to modulate angiogenesis using the rat aortic ring assay (RARA). For this assay, male Sprague-Dawley rats weighing 125-150 grams were sacrificed by an intramuscular followed by a cardiac injection of catemine, submerged in ethanol, and transferred to a sterile culture hood for dissection. Following a mid-abdominal incision extending to the thorax, the ribs were resected to expose the thoracic cavity. The heart and lungs were moved aside to locate the thoracic aorta. The segment of the aorta distal to the aortic arch and extending to the diaphragm was removed and placed immediately in MCDB-131 medium containing 4 mM L-glutamine and 50 µg/ml gentamycin (buffer A). Buffer A was gently pipeted through the aorta with a pipetman to remove blood. The fibrous tissue was surrounding the aorta was carefully removed and the aorta was sliced into 1-2 mm sections with sterile razor blades. The sections were collected in 50 ml conical tubes and washed five times with fresh buffer A.

20 Twenty four-well culture plates were coated with 0.2 ml autoclaved 1.5% agarose in buffer A. After the agarose solidified, the wells were filled with 1 ml buffer A. One aortic segment was submerged in each well and incubated in a 35.5 °C/5% CO₂ humidified incubator for 10 days. Buffer A was replaced every 2 days. The aortic segments were collected in 50 ml conical tubes and washed five times with fresh buffer A.

25 One milliliter of autoclaved 1.5% agarose in buffer A was added to each well of a twenty four-well culture plate and allowed to solidify. Agarose wells were cut from the solidified agarose with ethanol-sterilized cork borers and placed in twelve-well culture plates. Collagen solution was prepared by mixing 12.8 ml Collaborative Biomedical Products rat tail type I collagen with 2 ml 7.5X Dulbecco's Modified Eagle's Medium adjusted to pH 7 with approximately 0.2 ml 1N NaOH. One hundred microliters of collagen solution was added to the base of the agarose wells and allowed to set in a 35.5 °C/5% CO₂ humidified incubator. Three hundred microliters of collagen solution supplemented with 0.5 nM bFGF and the desired concentration of test compound was added and an aortic segment was submerged within the upper collagen layer. Test compounds were initially dissolved in ethanol at a stock concentration of 0.1 mM, and subsequently diluted in buffer A and finally collagen solution to obtain concentrations from 0.1 nM to 100 nM. After the collagen set, the agarose wells were surrounded by 1.5 ml buffer A supplemented with 0.5 nM bFGF and the desired concentration of test angiogenesis inhibitor compound. The buffer surrounding the

wells was replaced every 48 hours. After seven days, the vessels protruding from the aortic rings were counted under a microscope.

The results of this assay, set forth herein in Figure 1, demonstrate that the angiogenesis inhibitor compounds of the invention have excellent angiogenesis inhibitory activity (much better compared to the angiogenesis inhibitory activity of TNP-470).

Example 28

The plasma stability of the angiogenesis inhibitor compounds of the invention was also tested in human plasma at 37°C (at a 20 µM concentration of the angiogenesis inhibitor compound). The matrix minus the test angiogenesis inhibitor compound was equilibrated at 37°C for 3 minutes. The assay was initiated by addition of the test compound, and the reaction mixture was incubated in a shaking water bath at 37°C. Aliquots (100µL) were withdrawn in triplicate at 0, 60, and 120 minutes and combined with 200µL of ice cold acetonitrile to terminate the reaction. The supernatant was analysed by Liquid Chromatography/Mass spectrometry (LC/MS) using a four point standard curve to obtain percent recovery.

The parent compound remaining (relative to time 0) in the incubation mixture was plotted as a function of time; a first order exponential equation was fit to the observed data, and the elimination half-lives associated with the disappearance of the test angiogenesis inhibitor compound were determined. Results are shown below in Table VI.

Table VI

Test Compound	Percent Recovery (Remaining)						T _{1/2} (min)
	0 min	15 min	30 min	60 min	90 min	120 min	
5	100	108	103	97	129	107	≥500
20	110	114	114	120	106	103	≥500

25

The stability of the angiogenesis inhibitor compounds in liver microsomes was also tested. The test angiogenesis inhibitor compound at a concentration of 3µM was incubated with male rat liver microsomes (0.5 mg/mL protein) in the presence of a NADPH regenerating system (1 mM NADP, 10 mM G6P, 1 U/mL G6PdH2, 3 mM MgCl₂) in 100mM Phosphate buffer at pH 7.4 at 37°C. Incubations were performed in the presence and absence of 1µM epoxide hydrolase (EH) inhibitor N-cyclohexyl-N'-(3-phenylpropyl) urea (M. Morisseau *et al.* (1999) *PNAS* 96:8849-54) and quantitative analysis performed using LC/MS/MS.

The parent compound remaining (relative to time 0) in the incubation mixture was plotted as a function of time; a first order exponential equation was fit to the observed data, and the elimination half-lives associated with the disappearance of the test compound were determined. Results are shown below in Table VII.

Table VII

Test Compound	T _{1/2} (without EH inhibitor) mins	T _{1/2} (with EH inhibitor) mins
TNP-470	> 1.0	> 1.0
5	11.9 ± 1	12.1 ± 1

Example 29

The efficacy of the compound described above in example 5 was evaluated using a collagen-induced arthritis model. Briefly, syngeneic 8-week-old female Louvain rats were immunized intradermally under ether anesthesia on Day 0 with 0.5 mg chick collagen II (Genzyme, Boston, MA) solubilized in 0.1M acetic acid and emulsified in Incomplete Freund's Adjuvant (Difco, Detroit, MI). Using this method, more than 90% of rats typically develop synovitis in both hind limbs 10-14 days post immunization. Rats demonstrating definite arthritis (47 total) were entered into the study and randomly assigned to one of four protocols. The compound of example 5 (hereinafter "compound 5") was dissolved in a combination of ethanol and distilled water. The rats were dosed commencing on day 10 post immunization as follows: (1) control group: vehicle only daily via intravenous injection (iv) (12 rats), (2) high dose group: 30 mg/kg compound 5 iv every other day (12 rats), (3) low dose group: 15 mg/kg compound 5 iv every day (12 rats) and (4) oral group: 100 mg/kg compound 5 every other day by gavage (11 rats).

Daily clinical evaluations of each paw were graded on an integer scale ranging from 0 to 4, where a score of 0 indicates a normal, unaffected limb, and 4 indicates severe joint destruction. The clinical score is the sum of the four limb scores, and has a maximum possible value of 16. Collagen induced arthritis typically develops only in the hind limbs; therefore, a score of 6 to 8 represents severe arthritis.

Radiologic evaluations of the hind limbs were performed by an investigator blinded to the treatment protocol. The score assigned to each limb was based on (1) the degree of soft tissue swelling, (2) narrowing of the space between joints, (3) degree of periosteal new bone formation and (4) the presence of erosions or ankylosis. Each limb was scored on a scale of 1 to 3, with 1 representing a normal limb and 3 representing severe joint destruction. Each rat, thus, had a maximum possible radiographic score of 6 (sum of both hind limbs).

The results of this study are set forth in Figure 2, which presents the mean daily clinical score for each of the four groups of rats over the 19 day period following induction of clinical arthritis (day 10). The results indicate that each treatment group received a significant therapeutic benefit from compound 5. Consistent with these observations, the mean radiological evaluation scores were 2.2 for the control group, 0.6 for the high dose group, 0.6 for the low dose group and 1.6 for the oral group. Thus, each group treated with compound 5 received a significant benefit with regard to joint destruction compared to the control group.

10

Equivalents

Those skilled in the art will recognize, or be able to ascertain using no more than routine experimentation, many equivalents to the specific embodiments of the invention described herein. Such equivalents are intended to be encompassed by the following

5 claims.